

Purification, Determination Molecular Weight and Study Kinetic Properties of G6PD from Diabetes Patient

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Abstract

This study was conducted to purification G6PD enzyme from diabetic patients by using simple and cheap method the technique gel filtration on Sephadex G100 and determine molecular weight of enzyme and compare it with true molecular weight of enzyme and determine kinetic constant (Km, Vmax) and study the effect of temperature and substrate and pH and known the best condition to give optimum work of enzyme. study contain (60) patients with diabetes and (60) control Glucose and activity of G6PD were measured and the enzyme precipitated by Ammonium Sulfate with concentration (75%) and purification enzyme gel filtration on Sephadex G-100 with dimensions (1.5 × 30) cm and using the buffer solution from (Tris-HCl) at pH 8.2 to isolate the enzyme and determine molecular weight with same method. Specific activity was calculated (21.5 UI/mg), total activity (706.8 UI), number of purification (3.45) enzyme yield (23.188%) and enzyme activity (17.67 UI/ml). and the molecular weight was calculated with using same technique (57.82) kD. Effect of increased concentration of substrate on enzyme activity and found the activity increase with increase substrate and amount constant level not change however increase of concentration of substrate when drawing relation between activity and concentration of substrate format appear exchange excess and after study effect of pH found the optimum value (8.4) and study effect of temperature on activity found (38 C) the optimum temperature. Study of kinetic constant was done and the and the Michaelis-Menten (Km) value was (3.8 mM) and Vmax value (8 IU/ml).

Keywords: G6PD; Diabetes mellitus; Purification; Molecular weight

Introduction

Diabetes mellitus is a metabolic disease clinically and genetically heterogeneous group characterized by hyperglycemia due to defects in insulin metabolism. If the hyperglycemia of diabetes is not managed properly, it causes long-term damage, dysfunction, and failure of different organism [1]. Notably the eyes, kidneys, nerves, heart, and blood diabetes mellitus is a multifactorial disease resulting from interaction of both genetic and environmental It has been stated that oxidative stress and impaired release of nitric oxide may be the contributory factors in the pathogenesis of diabetes [2-4].

One of the main causes of diabetes is functional causes such as pancreatic disorders, this occurs when the pancreas is infected with tumors (benign and malignant), internal bleeding or when the pancreas is removed leading to an absolute inability to secrete insulin to lead to diabetes induced by excessive use of thiazide diuretics, anti-inflammatory drugs and antiviral [2]. Genetic causes Individuals with parents who have diabetes or individuals from a family with a family history of the disease are more likely to develop diabetes than others obesity is a cause of diabetes, Obese people who store high amounts of fat in the abdomen are more likely to develop diabetes [5]. Those who accumulate fat in the limbs and increase the amount of fats affect the blood sugar level because it is one of the main factors that cause insulin resistance of cells by reducing the sensitivity of insulin receptors on the surface of target cells [4]. One of the other reasons is also lack of physical activity as it affects the increased incidence of diabetes in some cases, emotional emotions have an effect on diabetes, such as anxiety, fear or sudden shocks, and susceptibility to disease [6]. Viral infections have a major role in the development of type 1 diabetes, Self-infection due to viral infection or by the destruction of beta cells in the pancreas [7] (Table 1).

Diagnosis of diabetes

Clinical diagnosis

Patients with diabetes of all types have the following symptoms: Polyuria, polydipsia, lethargy, boils, slow healing wounds, frequent infections persist for a long time. Patients with Type I diabetes suffer from weight loss dehydration, ketonuria, and hyperventilation [6]. Symptoms of type 1 diabetes tend to be short-term, whereas patients with type 2 diabetes tend to have chronic symptoms with longer duration of symptoms [5]. This is a significant difference between the two. Lack of secretion of insulin also causes excessive metabolism of free fatty acids, and this leads to confusion in fat metabolism [7].

Laboratory diagnosis: High blood sugar hyperglycemia and the emergence of glucose in the blood glucose is a distinctive phenomenon of diabetes, so some tests are used to diagnose diabetes, including: Fasting Plasma Glucose (FPG), Random plasma Glucose (RPG), Oral glucose Tolerance test (OGTT), Glycosylated Haemoglobin (HbA_{1c}), Glucose in Urine, Ketone body in Blood or Urine [8,9].

Glucose-6-Phosphate Dehydrogenase (G6PD)

Enzyme: Are vital catalysts that accelerate the rate of chemical reactions. They have a high molecular weight protein structure. Like other proteins, the enzyme is composed of a combination of a large

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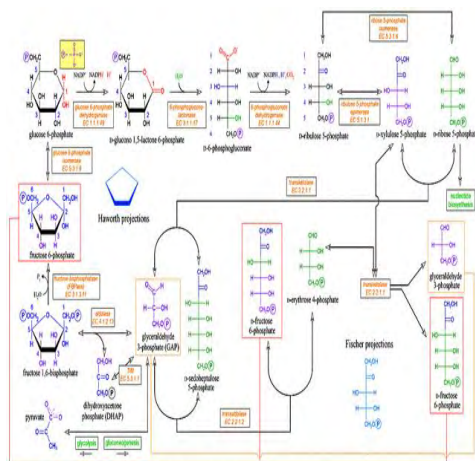
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number of amino acids that have one or more polypeptides. Is a three-dimensional form of the protein? Amino acids are found in these sequences according to a particular sequence of each enzyme, leading to a specific vacuum structure that enables the enzyme to accelerate its own reaction [10-12].

Glucose-6-Phosphate Dehydrogenase (G6PD ((Oxidoreductase, EC1.1.1.49) (G6PD) is one of the cytoplasmic enzymes is spread throughout the body, especially in the red blood cells, which is one of the most important enzymes of the egg, as it is the main enzyme and the key to the Pentose phosphate pathway [13]. It stimulates the oxidation process of the glucose-6-phosphate (G-6-P) (NADP) and to convert it to an effective reduced form (NADP) to preserve the life-producing pathways of several important substances, particularly in red blood cells because they have no other source of production (NADP) to preserve the life-producing pathways of several important substances, particularly in red blood cells because they do not have another source of NADPH production. NADPH product produced by G6PD is complementary to the reduced triglyceride enzyme (GRG), which converts and converts oxidative glutathione (GSSG) (GSH) [7], which protects human red blood cells from partial and planned (1), shows the pathway of pentose phosphate sugar and rule of G6PD in reactions of pathway.



Planning (1) Pentose phosphate pathway [8].

The enzyme was first discovered by the scientists Warburg (Christian) in 1931 in the red blood cells of the horse and since then studies and research have been conducted to extract and purify the enzyme from various sources [9,14-16].

There are other important reasons to study as the change in the effectiveness of this enzyme G6PD enzyme in the body's various tissues is linked to many diseases in humans and these diseases is the disease of jaundice in children and hemolytic anemia [17]. G6PD deficiency is a disease that is prevalent in different parts of the world. The number of infected people is 400 million males, females, neonates and other ages, according to the World Health Organization (WHO) report [1]. More than 442 types of enzyme (G6PD variants) have been identified using a large number of biological techniques, including molecular analytical methods, which identify genetic mutations that occur in the genes responsible for the biological processing of different types of enzyme [10]. Gene found that the enzyme gene was found to be carried on the sex chromosome (X) [12]. The shortening of enzyme efficacy is associated with genetic and hereditary disorders [11]. This shortage is widespread in the world, especially in the Mediterranean region Patients with this

type of hemorrhagic deficiency are generally affected by certain drugs and foods in the case of a paroxysmal or neonatal jaundice found in natural erythrocytes, enzymatic activity decreases with age [11]. Many mutations of this enzymatic deficiency are widespread in the world and by geographic location. Moreover, genetic defects and age can lead to Enzymatic Deficiency An enzyme deficiency leads to the production of some red blood cell anemia due to exposure to certain chemical agents or certain infections and wounds [12].

Classification

Numerous G6PD variants have been described These have been classified by the World Health Organization according to the magnitude of the enzyme deficiency and the severity of hemolysis. This classification gives some approximation of the magnitude of hemolysis an individual might incur in the setting of an oxidative stress. Only class I, II, and III are of clinical significance.

Class I – Class I variants have severe enzyme deficiency (<10 percent of normal) relation with chronic hemolytic anemia.

Class II – Class II variants also have severe enzyme deficiency (<10 percent of normal), but there is usually only intermittent hemolysis, typically on exposure to oxidant stress such as fava bean exposure or ingestion of certain drugs and the classic example is (G6PD Mediterranean).

Class III – Class III variants have moderate enzyme deficiency (10 to 60 percent of normal) with intermittent hemolysis, typically associated with significant oxidant stress the classic example is (G6PD A).

Class IV – Class IV variants have no enzyme deficiency or hemolysis the wild-type (normal) enzyme is considered a class IV variant, as are numerous other genetic changes that do not alter levels of the enzyme and these variants are of no clinical significance.

Class V – Class V variants have increased enzyme activity (more than twice normal). These are typically uncovered during testing for G6PD deficiency and they are of no clinical significance [13].

Clinical Significance of the Enzyme

Acute hemolytic anemia

Some individuals with G6PD deficiency have acute hemolytic anemia at the site of the wound when some medications, acute diseases, and certain foods are taken [15].

Neonatal jaundice

Anemia and jaundice are most often observed in newborns in individuals with severe enzyme deficiency [18-21].

Neutrophil dysfunction

The enzyme is used in addition to red blood cells in white cells to reduce the oxidizing factors. Some people with severe enzyme deficiency suffer from a defect in the function of white blood cells, which causes weak respiratory resistance to diseases and also weakens the presence of beneficial bacteria in the body [14].

Diabetic mellitus-induced hemolysis

In people with an enzyme deficiency, hemolyticcysis starts with an increase in ketone content in diabetics and has the lowest levels when blood glucose levels are abnormal in diabetics [22]. Studies have indicated that high sugar leads to the deposition of decomposed blood

in patients with deficiency [15].

Relationship Between (G6PD) Enzyme and Diabetes

An epidemiological study from suggested a positive correlation between diabetes and deficiency [16]. (G6PD) has conducted a study on Indian society being the most potential to give this relationship Serum samples were collected for healthy people and of both sexes. A higher incidence among Indians provided an excellent opportunity to study the possible association of G6PD deficiency in diabetes mellitus [5]. G6 PD deficiency is one of the common enzymopathy in human being affecting about 400 million people worldwide. It is suggested that there may be a positive association of G6PD deficiency with diabetes mellitus. Although G6PD deficiency is not uncommon in our country but there is scarcity of data on this regard especially on diabetes. Therefore, this study was undertaken to observe the G6PD status in patients with type 2 diabetes mellitus in order to explore the role of this enzyme deficiency as one of the risk factor for diabetes mellitus [7]. A positive association of G6PD deficiency with diabetes mellitus Although G6PD deficiency is not uncommon in our country but there is scarcity of data on this regard especially on diabetes. Therefore, this study was undertaken to observe the G6PD status in patients with type 2 diabetes mellitus in order to explore the role of this enzyme deficiency as one of the risk factor for diabetes mellitus [7].

Materials and Methods

Collection of sample

The total number of these samples was (60) samples, serum samples were collected for people with diabetes and both sexes. They reached (60) satisfactory samples of both types of diabetes (type I and type II) Diagnosis of the disease using a blood glucose test.

Blood was drawn from the vein using a 5 ml plastic syringe with one use. The blood was placed in clean, sterile plastic tubes free of anticoagulant EDTA. And left to coagulate at room temperature. The blood serum was then separated from the centrifuged portion of the centrifuge and at a velocity of 5000 G for 15 minutes to ensure adequate serum red blood cell extraction, Micro pipette. The effectiveness of the enzyme was measured directly and the study was done outside the body (*in vitro*) [23-28].

Diagnosis kits

The kits Clarifiers in Table 2 were used in procedures of this study.

Estimation of Biochemical Parameters in Blood Serum

Estimation of glucose concentration in serum

Principle: glucose level in serum was measured by using (kit Aflu Italia) depending on enzyme method that stated on Trinder reaction [29].

Determination of total protein in serum

Total protein level in serum was measured by using (kit Aflu Italia) depending on enzyme method [30].

Determination of G6PD activity in serum

G6PD activity in serum was measured by using Biolabo kit according this equation [31]:

Where; A=Absorbance; V=enzyme volume in ml; 6.22=absorbance confection of (NADPH) in length 340 nm.

Separation and Purification of the Enzyme G6PDH from Serum Diabetes Patients

G6PDH was purified from the serum of diabetic patients using the following steps:

Addition ammonium sulphate

Serum proteins were precipitated using gradual concentrations of ammonium sulphate until 75%. 3.75 gm of ammonium sulphate was added to 5 ml of serum during 60-45 minutes by placing the serum in a snow bath with constant stirring, and then dissolved Deposition using 4 ml of 1M Tris-HCl regulated solution (pH=7.8) [32].

Dialysis

Which is one of the most important methods used in the purification of enzymes and the oldest, and the goal is to remove the remaining ammonium sulphate added to the deposition of proteins by placing the dissolved protein in the above step in the membrane bag dialysis bag after measuring the effectiveness of the enzyme G6PD and protein concentration, and immerses the bag in the solution [33-35]. 1 M Tris-HCl pH 7.8 The regulator solution was changed from time to time for 16 hours. This step was performed at (4°C ± 1°C) to maintain the efficacy of G6PD. After the membrane separation process After the membrane separation process, G6PD and protein concentration were measured.

Gel filtration chromatography

Gel Filtration technology is one of the most important techniques in the field of biochemistry, one of the methods used in the separation of compounds depending on the size of their molecules and their molecular weights. The proteins with large molecular weights are not carried out through the gel but move outside the gel layer with the solvent that is removed in sequence. This solvent is often distilled water or a dilute regulator. Therefore, the large particles first filter during the separation. Small molecules can penetrate the gel granules finally. The fraction of the gel filtration is collected using the Fraction Collector. The volume of distilled water or solution is calculated to displace each

Year 2030	Year 2010	Country
2.009.000	968.000	Iraq
6.725.000	2.873.000	Egypt
680.000	315.000	Jordan
2.523.000	895.000	Saudi Arabia
99.000	38.000	Bahrain
378.000	149.000	Lebanon
1.138.000	437.000	Morocco

Table 1: Statistical represents the number of patients with diabetes in the Arab States in 2010, and it is expected in 2030 [4].

1	Glucose 6-Phosphate dehydrogenase (G6PDH) kit	Biolabo- France
2	Total protein kit	Aflu-Italia
3	Glucose kit	Aflu-Italia

Table 2: The kits and its source.

protein from the separation column, and the protein is isolated by reading the absorption at 280 nm [17,36-38].

The gel filtration technology is also used as a method for estimating the approximate molecular weight of protein by drawing a graph showing the relationship between the size of the elution and the known molecular weight. The protein with the molecular weight is then passed through the separation column and calculated. The size of the Rogan accurately and in comparison with the known molecular weights can estimate the approximate molecular weight of the unknown protein [18,19].

Used solutions

buffer solution 0.1M Tris-HCl pH 7.8. Prepare to dissolve 15.76 gm of Tris-HCl per liter of distilled water and adjust pH at 7.8.

Sephadex G100 Prepare to dissolve 2.5 gm of the Sephadex G100 column filler in 200 ml of the 0.1 M Tris-HCl pH solution 7.2 and leave the solution for 28-24 hours at 4°C. During this time, the solution was changed several times to remove the soft minutes from the solution Because its presence reduces the velocity of the flow of the liquid solution through the column [39].

Sodium chloride solution at 500 mM concentration Prepare 29.25 g of NaCl per liter of 0.1 M Tris-HCl pH 7.8 solution.

Procedure

Use a glass column with a diameter of 1.5 cm and a length of 30 cm. A small amount of glass wool is placed at the end of the column to prevent the gel particles from leaking out of the column. The gel solution is slowly and homogeneously poured into the column to prevent air bubbles from forming. (11 cm), wash the column with sufficient amounts of 0.1M Tris-HCl pH solution until a flow velocity of 2.5 ml/min was obtained [20].

Add 5 ml of enzyme after membrane separation slowly over the surface of the G100 Sephadex gel and leave for 5 minutes to soak in the gel column.

The process of separation was started using 150 ml of the structured solution containing 500 ml of NaCl, collecting 5 ml per part.

After collecting the extracting parts of the separation column, the efficacy of the G6PD enzyme was evaluated by paragraph and protein concentration by the method (kit Aflu).

Kinetics of G6PD

The kinetics of G6PD were studied after its separation and partially purified from the serum of diabetic patients by gel filtration. These included:

Effect of Glucose 6-phosphate concentration (G6P)

The effect of different concentrations of G6P on the activity of G6PD was studied by using different concentrations (0.6, 0.0512, 0.0256, 0.0128, 0.064, 0.048, 0.024, 0.012) M to determine the effect of the concentration of substrate on the work of the enzyme G6PD, G6PD reaction (G6PD kit Biolabo), and plotting the relationship between the reaction rate and the concentration of substrate to determine that the enzyme is subject to the Michaelis-Menten equation. The Km values were obtained using the Linover-Burke graphical method, which links the inverse values of both velocity and G6P concentration ($1/(S)$ vs. $1/V$) [40].

Optimized pH mapping

The pH effect of the regulated solution (0.1M Tris-HCl pH 7.8) was studied at the velocity of the G6PD reaction. Different pH solutions (11, 10, 9, 8, 7.6) were used with G6P at 0.6 mM and 37°C (G6PD kit Biolabo), and by plotting the relationship between reaction velocity and pH, the optimal pH was identified.

Effect of temperature

G6PD kit Biolabo was used to measure the effectiveness of G6PD. The reaction was conducted at different temperatures (57, 47, 37, 27, 17 and 7) with the regulated solution (0.1M Tris - HCl pH 7.8) Basically (G6P) 0.6 mM, and then painted the relationship between the reaction velocity and the temperature to find out the optimal temperature of the reaction [41].

Results and Discussion

Purification of enzyme

Precipitation by ammonium sulphate: The basic principle of the method is to equalize the charges on the surface of the protein and the degradation of the water layer surrounding the protein and reduce the degree of watering and reduce the solubility of the protein and sedimentation [23]. Add the stages to get rid of some of the protein content with the enzymatic extract [24]. The result in Table 3 showing the efficiency of quality (10.77 units/mg) with the number of times of purification (1.73) and the yield (35.02) during the satisfaction rate of ammonium sulfate sulfate estimated (75%). The results differed with the studies of enzyme extraction. In a study involving red blood cell extraction, the specific activity was 1.251 (units/mg) with 121.5 purification number and 53.8% enzymatic yield when using ammonium sulphate salt at 35-65% [25]. Another study showed that the specific efficacy was 0.37 mg/ml and the number of purification times was 39.79 times and the yield of 79.18 units/mg after adding the salt with saturation concentration (40-60%) of the red blood cells of the geese [26]. In the Penicillium duPonti fungi, the salt was used by saturation (45-60%), giving the result a quality efficacy of (1.04 units/mg) and the number of times the purification of 8.67 times and proceeds 63.4% [33].

Gel filtration: The different methods used to purify the enzyme from bacterial, fungal, plant, or animal sources were obtained in obtaining high purity of the enzyme, during this research the use of gel filtration technique with the Sephadex G-100 was the result of the specific efficacy (21.5 units/mg), total activity (706.8 units), purification number 3.45, enzymatic yield (23.188%) and enzymatic efficacy (17.67 units/ml). Results were obtained with other studies of enzyme purification. In one study, using the Sephacryl-S200 column to purify the enzyme from rat liver it was found that the specific efficacy was 24.75 units/mg and the total efficiency was 198 units with 6.17 times purification and an enzyme yield of 60.57% [28]. In a study that included several steps, first transfer the concentrated enzymatic extract with ammonium sulphate salt on the calcium phosphate column and then transfer the extract to the ion exchanger DEAE-Cellulose and then the gel filtration column Bio-Gel A-150 these steps gave a quality effect of 470 units/mg with a frequency of 2.42 times and an enzyme yield of 10% [29]. A study of the purified enzyme from calf tissue using the gel filter column Sephadex G-25 an enzyme yield of 91% with 450 times purification. Another study included enzyme purification from Coriander Leaves The first two-step purification was performed with ammonium sulphate deposition and the use of the Sephadex G-200 gel. The enzymatic activity was 1.82 units/mg and the enzymatic yield was

26.4% and the number of purification times was 74 [30]. Also using gelatin filtration of the Sephadex G-200 column and concentration of sulphate salts prior to filtration, the results included (326 unit/mg) the enzymatic yield was 19.9% and the frequency of purification was 2.5 when purifying the enzyme from the pituitary gland of cow [31].

In another study to *Saccharomyces cerevisiae* include using saturated concentration of ammonium sulphate salt (40-80) and gel filtration column on Sephacryl S-200 gave specific activity 65.68 unit/mg and the enzymatic yield was 20.62% and the number of purification times was 2.94 [29].

Determination of Molecular Weight in Gelatin Filtration Technology

The researcher's method [26] was based on the gel filter method in estimating the approximate molecular weight of the G6PD enzyme from the protein package (18) which showed the highest concentration of the protein. and passed a number of known molecular weight indicated in Table 4 and the molecular weight ranges between (20000000-13700) Dalton for the purpose of specifying the characteristics of the column. In terms of internal volume (Elution Volume V_e) for each material as well as the free or empty size of the granules (Void Volume V_o), which was estimated from the standard curve of blue dextran at 3 mL per part. The V_o value was equal to 33 mL and the recovery volume of the standard V_e proteins to the volume of the recovery of dextran blue V_o , represented by the V_e/V_o relationship [28]. As shown in Table 4 (Sephadex G-100), molecular weight and recovery volume The Elution Volume of each material versus its molecular weight logarithm shows the appearance of a straight line in which the approximate molecular weight of the protein packet separated by the gel filtration technique is shown in Figure 1. The recovery volume of the package (18 ml) To approximate molecular weight (Figure 2), the approximate molecular weight of the G6PD is (57.82) kDa for the enzymatic extract using gel filtration technique.

We see these results contrasted with a range of studies and studies conducted to estimate the molecular weight of the enzyme from its various sources [34]. The molecular weight was 133 kDa for the purified enzyme from pig liver using ammonium sulphate and Sephadex G-200 for the bilateral body the single molecule had a molecular weight of 67.50 and other studies showed that the molecular weight of the enzyme was purified from the liver of the mice, and the pituitary gland for cows

had a molecular weight of 64 kDa [32]. The molecular weight of the purified enzyme from human red blood cells was estimated to be 43 kDa [34]. and in another study using gelatin filtration with the Sephadex G150 the molecular weight was (40 KDa) from Bean plant [35]. The molecular weight of the purified enzyme from diabetics was within the range mentioned [41], ranging from 22-58 KDa. Due to the difference in the scientific basis in the different methods used in the Dalton of the enzyme purified from yeast *Saccharomyces cerevisiae* Purification of enzymes The molecular weights vary from one study to another [40]. The difference is also due to the length of the purification steps of some of the different studies which may lead to the enzyme breaking through these long stages and therefore the resulting molecular weights are less than the real G6PD Study of Enzyme kinetics [42-44].

G6PD Study of Enzyme Kinetics

Effect of the pH

pH has a significant effect on the enzyme's effectiveness for controlling ionization Ionic aggregates at the active site of the enzyme The optimal pH of enzyme stability is an important characteristic of enzymes [38]. The results of the kinetic study of the enzyme showed that the optimal pH of the enzymatic extract was in the range (8-8.4) as shown in Figure 3. The optimal basis for the stability of the enzyme was (8-9) Extreme pH values affect substrate and the ionic state of the enzyme and lead to enzyme protein mutagenesis by altering the enzyme body [23]. In a study of [43] people with enzymatic deficiency and a group of healthy patients, pH was (8.5-7) as in Figure 4.

Filter column the gel filtration column using the Sephadex gel and the solution of regulating the hydrochloride gear with different hydrogen numbers ranging from 5-11, respectively.

Effect of temperature on enzyme activity

The optimum temperature of the activity, the highest temperature, where the rate of enzymatic reaction rate is maximal, while the enzyme is highly effective, and is affected by pH and other factors [39]. The optimum temperature of the enzyme activity when the pH was confirmed and the concentration of Substrate was (37-38°C). The results varied with the studies carried out, including a study on the enzymatic extract extracted from coriander leaves (30C) [31]. and another study indicated that the optimum degree of the enzyme purified from bacteria *Azotobacter* and human placenta is (50C) [36].

Step of purification	Elute (ml)	Activity (IU/ml)	Protein conc. mg/ml	Specific Activity (IU/mg)	Total Activity (IU)	Purification (fold)	Yield (%)	Total Protein conc. (mg)
Crud serum	5	30.48	24.5	6.22	3048	1	100	4.9
Ammonium sulphate (75%)	4	21.54	8	10.77	1077	1.73	35	2
Dialysis	3.5	19.1	6.37	10.49	1049	1.68	34.4	1.82
Gel filtration (Sephadex G-100)	3	17.67	2.46	21.5	706.8	3.45	23.188	0.82

Table 3: Steps of enzyme purification.

Standard protein	Molecular weight (Dalton)	Log Molecular weight	Number of fraction (ml)	Elution volume (V_e) in (ml)	V_e/V_o
Blue Dextran	2000000	6.3	11	33	1
BSA	67000	4.82	15	45	1.36
alpha-Amylase Enzyme	58000	4.7	20	60	1.3
ovalbumin	43000	4.63	24	72	2.18
Chymotrypsinogen	23000	4.39	27	81	2.45
Ribonuclease	13700	4.13	31	93	2.81

Table 4: Relation between molecular weight and elution volume to standard protein.

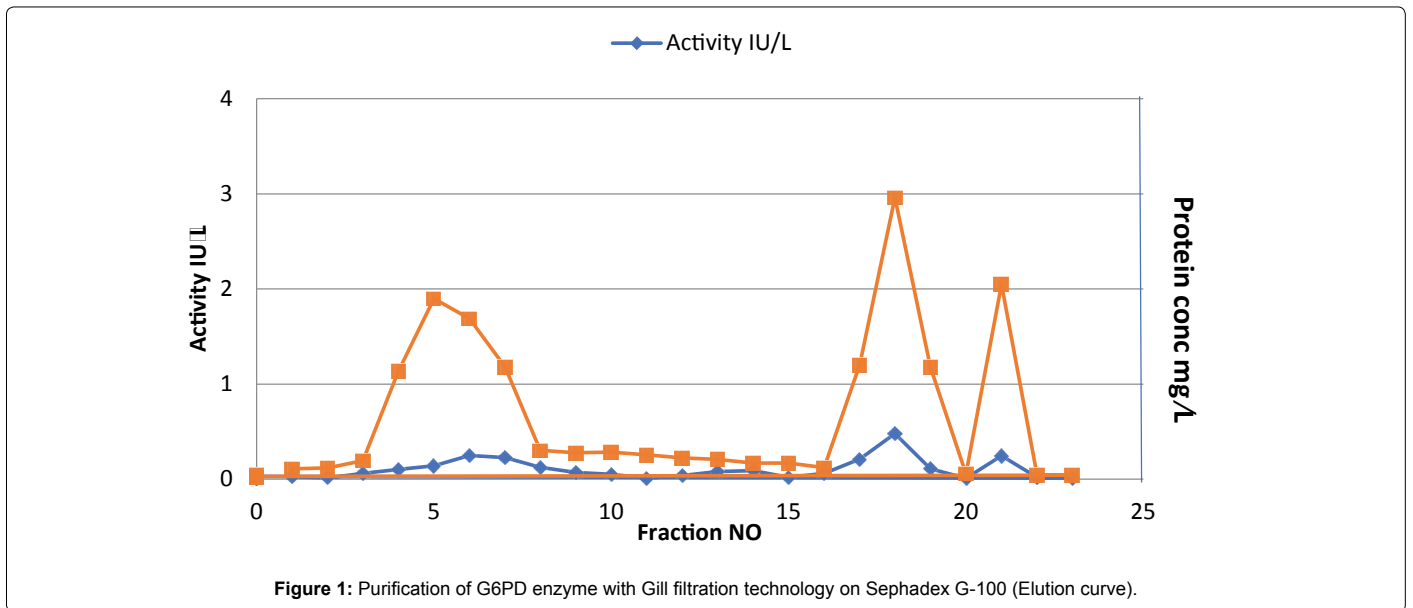


Figure 1: Purification of G6PD enzyme with Gill filtration technology on Sephadex G-100 (Elution curve).

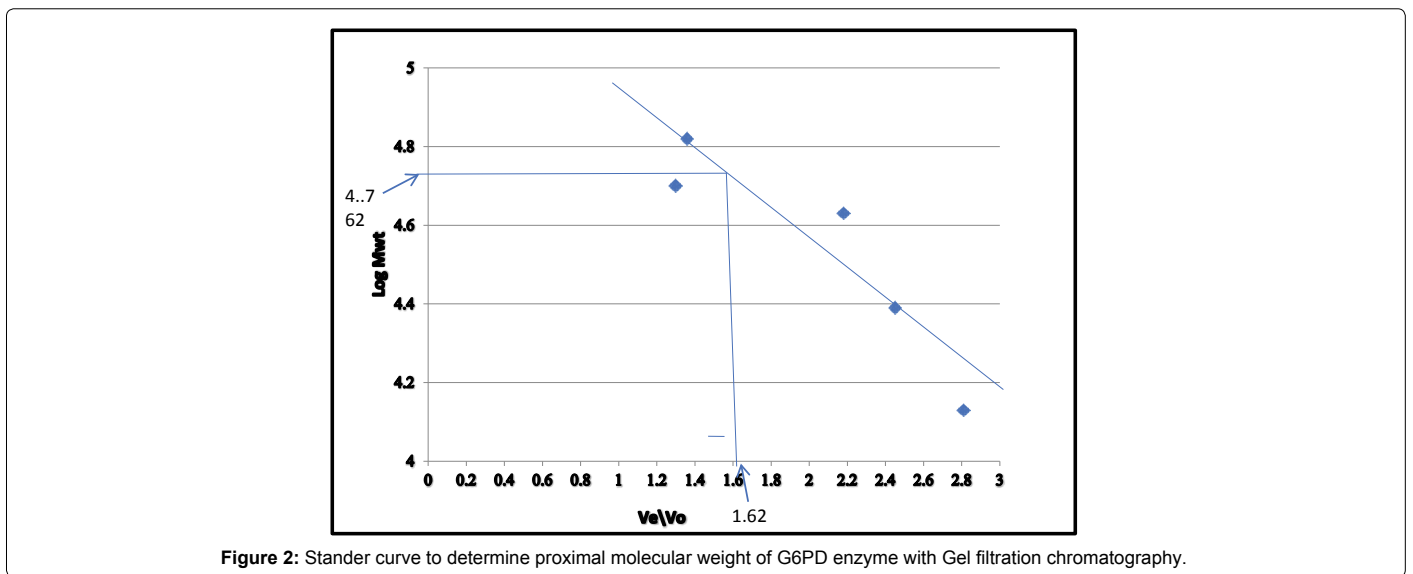


Figure 2: Stander curve to determine proximal molecular weight of G6PD enzyme with Gel filtration chromatography.

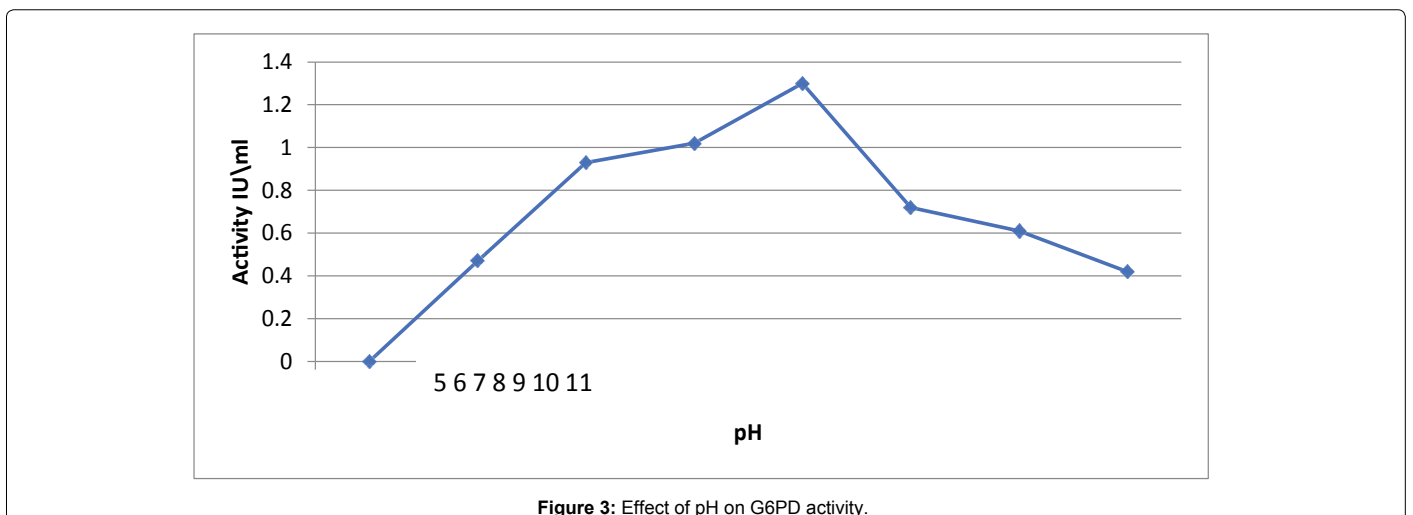


Figure 3: Effect of pH on G6PD activity.

In a study of the purified extract from the lamb marrow, the optimum grade was (38C) [37].

Effect of substrate concentration

The Michaelis-Menten constant (K_m) was defined as the affinity between the enzyme and substrate. The higher its value, the less the value of the substance is reduced. In order to stimulate the biological reactions and to determine the stability of enzymes and the effect of inhibitory and activating substances on enzymatic efficacy [40]. The results of the kinetic constants estimated for the enzymatic extract were shown in Figures 5 and 6. The constant value of the Michaelis-Menten of substrate (G6P) was (3.8mM) and the maximum velocity value V_{max} (8 UI/ml). The differences between all these studies were clear and almost natural as a result of the different sources of the enzyme which were cleared and the different methods In the Iraqi study to purify the enzyme from human blood, the value of Michaelis-Menten was found

in the substrate after the use of two purification methods (103 and 114 micro mole) and the maximum velocity (362 and 403 micromole/min/mg) respectively [41]. In another study on coriander leaves, the value of the Michaelis-Menten constant was 0.11 mmol and the maximum velocity was 0.038 units/ml. In a study to purify the enzyme from the pituitary gland for cows, the values of the Michaelis-Menten constant were 0.042 mmol and the maximum velocity was 9 units/ml [32]. In a WHO report for 2015, Michaelis Menten - purified from malaria patients is an average (30-50 micro mole) [1]. In a study on fungus *Penicillium duPonti* the Michaelis constant and the maximum velocity were respectively (0.43 mmol, 9 unit/mg) for the purified enzyme of the fungus [33]. The value of the Michaelis constant (K_m) from the human placenta was (0.4 mmol) and the maximum velocity was (8 unit/mg) [36]. study of the enzyme purification of the local isolation of yeast *Saccharomyces cerevisiae* showed that the value of the Michaelis-Menten (K_m) constant was (0.343 mmol) and the maximum velocity of substrate G6P (4.08 mmol/min) [45].

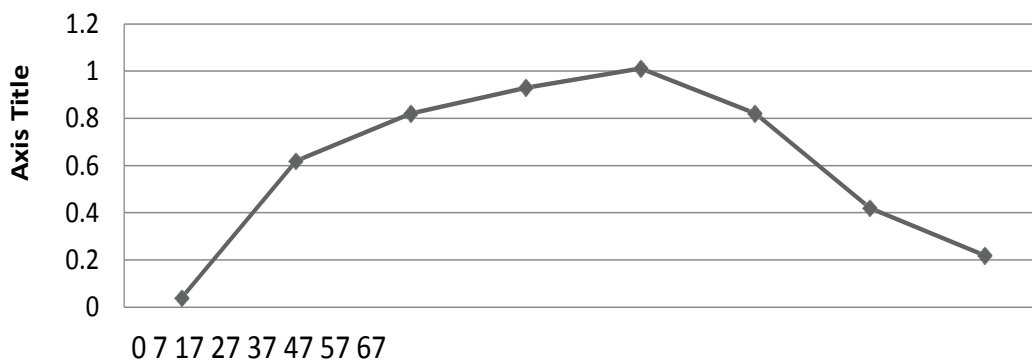


Figure 4: Effect of temperature on G6PD activity.

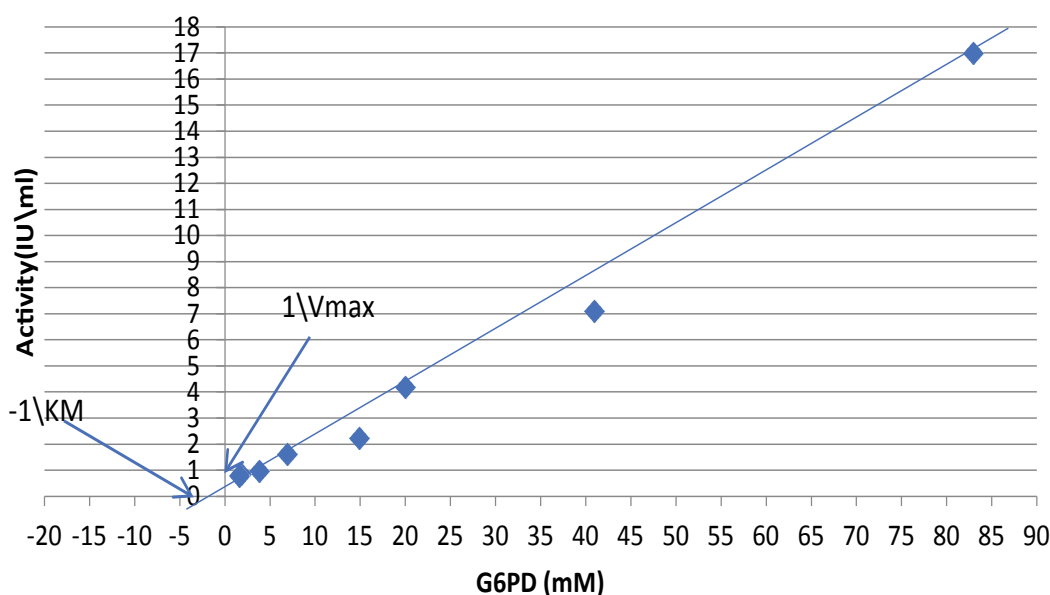


Figure 5: Lineweaver-Burk to calculate (K_m) and (V_{max}) to G6PD enzyme that purification from diabetes patient with Gill filtration on Sephadex G-100 towards substrate G6P with concentration between (0.6-0.0512 mM) with Coenzyme ($NADP^+=0.2$ Mm).

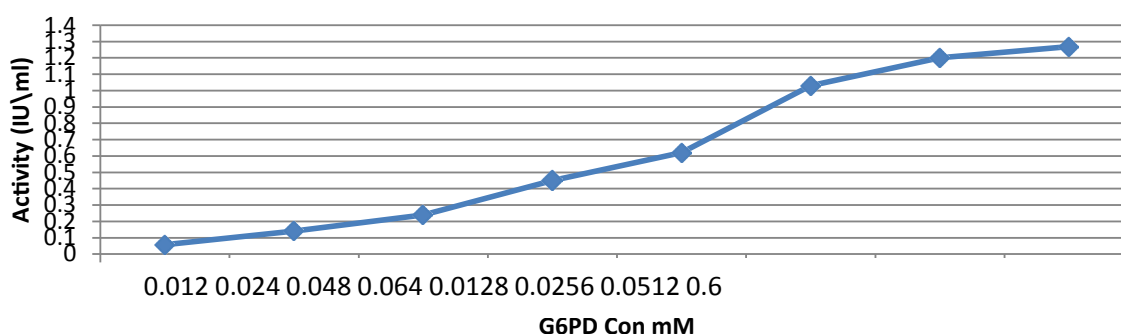


Figure 6: Effect of substrate concentration on enzyme activity.

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