

Isolation of Marine Bacteria From Visakhapatnam Coast For Degradation of Oil

Kiranmai MR¹ and Anima SD²

¹Environmental Sciences, GITAM Institute of Technology, GITAM University, Visakhapatnam, Andhra Pradesh, India

²Department of Chemistry, GITAM Institute of Science, GITAM University, Visakhapatnam, Andhra Pradesh, India

*Corresponding author: Kiranmai Reddy M, Environmental Sciences, GITAM Institute of Technology, GITAM University, Visakhapatnam, Andhra Pradesh, India, Tel: 9703868942; E-mail: kiranmai@gitam.edu

Rec date: March 08, 2017; Acc date: April 04, 2017; Pub date: April 06, 2017

Copyright: © 2017 Kiranmai MR, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Abstract

Marine waters are the receivers of many wastes from different sources such as food waste, raw material extraction, recreational use, waste disposal and transportation. In addition to this ship trafficking and industry releases enormous amounts of oil and grease to waters. The aim of the present study was to isolate bacterial strain from marine waters of Visakhapatnam coastal area in order to a strain suitable for the degradation of oil and grease. Many bacterial strains were isolated including *Aeromonas hydrophilia*, *Bacillus cereus*, *Bacillus subtilis*, *Enterobacter aerogenes*, *Escherichia coli*, *Proteus vulgaris*, *Pseudomonas aeruginosa*, *Salmonella typhimurium*, *Staphylococcus aureus*. The results showed that the degradation of oil and grease was possible by the application of extracted bacterial isolates. In our study, *Bacillus cereus* was identified as an efficient degrader of oil and grease compared to other isolates. This study helped to identify native marine bacteria from the polluted site which can be potentially used in biodegradation process of oil and grease.

Keywords: Marine waters; *Bacillus cereus*; Visakhapatnam

Introduction

Oceans are the sinks for the pollutants traveled through several sources such as rivers, streams, coastal restaurants, industries and inland transportation. Some of these sources are the contributors of oil wastes. As oil forms a layer on the surface of the ocean waters, it makes a hindrance of sunlight, oxygen for aquatic flora and fauna [1-3]. It is therefore of high interest to develop rapid and reliable tools for the analysis and monitoring of the fractions of oil or hydrocarbons. Physical and chemical methods like volatilization, photooxidation, chemical oxidation, and bioaccumulation [4] are rarely successful in rapid removal and cleaning up oil or hydrocarbons and also these methods are not safe and cost effective when compared to microbial methods. Crude oil sources are important energy resources used by industry and in our daily life. At the same time, crude oil products is a major pollutant of the environment [5]. Due to its complicated composition, it has the potential to elicit multiple types of toxic effects. Up to now there have been many reports concerning the role of marine microorganisms active in degradation of pollutant oil [6]. Oil products many contain molecules called hopanoids which were observed in bacterial cell wall [7,8] indicating biodegradation to some extent. Degradation of crude oil by naturally occurring bacterial population converts complex and hazardous substances into simpler and nontoxic forms which can be easily removed from the environment [1,9]. Some of research work related the degradation of hydrocarbons by microorganisms in different coastal regions [10,11], there are no available data on the degradation of hydrocarbons using microorganisms isolated at the region of the Visakhapatnam coast of Bay of Bengal. Moreover, there was no information about the distribution of marine microbes through the coastal line. The aim of the present study was to identify marine bacteria potentially useful for

the degradation of oil and also to assess the occurrence and the distribution of these marine bacteria.

Materials and Methods

Sample collection

Marine water samples were collected aseptically in a clean and sterilized container from three stations (station 1: ship trafficking, station 2: leakage of oil near harbour, station 3: restaurant oil waste release) covering about 1 km stretch through the harbour coast Visakhapatnam. The collected samples were immediately sent to the laboratory in an ice box for bacteriological examination [12] and analyzed physicochemical parameters (pH, temperature). 25 samples were collected at a stretch of 3 Km. Each sample was stored at 4°C prior to examination.

Isolation of marine bacteria

The pure strains of marine bacteria were isolated from the collected marine samples. 5.0 ml amounts of each marine sample were taken in a series of 250 ml Erlenmeyer flasks containing 45 ml of nutrient broth (Merck) and incubated at 35°C for 24 hours at 200 rpm in orbital shaker [13]. After incubation one loopful of each bacterial culture broth was inoculated on the nutrient agar plates and incubated in 35°C for 24 hours. The obtained bacterial colonies were sub-cultured on a fresh nutrient agar plats using similar procedures to get pure cultures.

Identification of bacteria

The obtained pure cultures were tested for gram positive and gram negative bacteria through Gram's staining. Then biochemical studies were conducted for the confirmation of bacterial species.

Biodegradation studies

Five ml of heavy machine cleaning oil was added to 100 ml of sterilized marine water in a series of 250 ml Erlenmeyer flasks and inoculated with 0.2 ml, 0.5 ml, 0.8 ml of the cultured pure strains respectively. The samples were examined on 3rd day, 5th day, 7th day, 9th day 11th day, 13th day, 15th day, and 17th day for the oil degradation. The samples were centrifuged, in which oil will be separated from water and the volume of the oil was noted.

Results and Discussion

In our study the bacterial strains were identified as *Aeromonas hydrophilia*, *Bacillus cereus*, *Bacillus subtilis*, *Enterobacter aerogenes*, *Escherichia coli*, *Proteus vulgaris*, *Pseudomonas aeruginosa*, *Salmonella typhimurium*, *Staphylococcus aureus*. All these species were identified based on the biochemical features showed in Table 1. One of the physicochemical parameter showed a significant role in the degradation abilities of bacteria.

S.No	Biochemical Analysis	<i>Bacillus cereus</i>	<i>Bacillus subtilis</i>	<i>Enterobacter aerogenes</i>	<i>Escherichia coli</i>	<i>Proteus vulgaris</i>	<i>Pseudomonas auregenosis</i>	<i>Salmonella typhimurium</i>	<i>Staphylococcus aureus</i>
1.	Starch Hydrolyss	-	+	-	+	-	-	-	-
2.	Lipid hydrolysis	+	+	-	+	+	-	-	-
3.	Casein Hydrolysis	+	+	-	+	+	-	-	-
4.	Hydrogen sulphide test	-	-	-	-	+	-	-	-
5.	Urease test	-	-	-	-	+	+	-	-
6.	MR-VP test	-	-	+	+	-	-	-	-
7.	Carbohydrate fermentation test	-	-	-	+	-	-	-	+
8.	Litmus milk test	-	-	-	-	+	+	-	-
9.	Catalase test	-	-	-	-	-	-	-	+

Table 1: Showing the biochemical analysis of extracted marine bacteria.

Influence of pH

pH played a very significant role in the biodegradation of bacteria on oil. In our study this was clearly noticed that the oil degradation was determined to happen at pH range of 6.5, 7.0, 7.5, and 8.0 respectively, where as insignificant degradation was observed at the pH range of 6.0, 5.5, 5.0 and 4.5 pointing towards the less survival of the cells. The degradation of oil depends on the presence of active bacteria in the sample. Ahmad et al. [13] stated that for an efficient bacterial bioremediation process minimum 10^5 CFU/ml of cell concentration is required. According to Nwuche and Oghonna [14] high oil concentration in low water availability and low pH decreases the bacterial activity. Similar conditions were identified in station 2 of the study area.

In case of samples collected from sampling stations 1 and 3 maximum degradation was achieved at pH 7.5 (Figure 1). The degradation of oil at different pH [7.0-8.0] increased or decreased. The degradation might be attributed to the variation of oil structures due to low or high pH values that would influence non-living degradation process according to Leahy and Colwell [1].

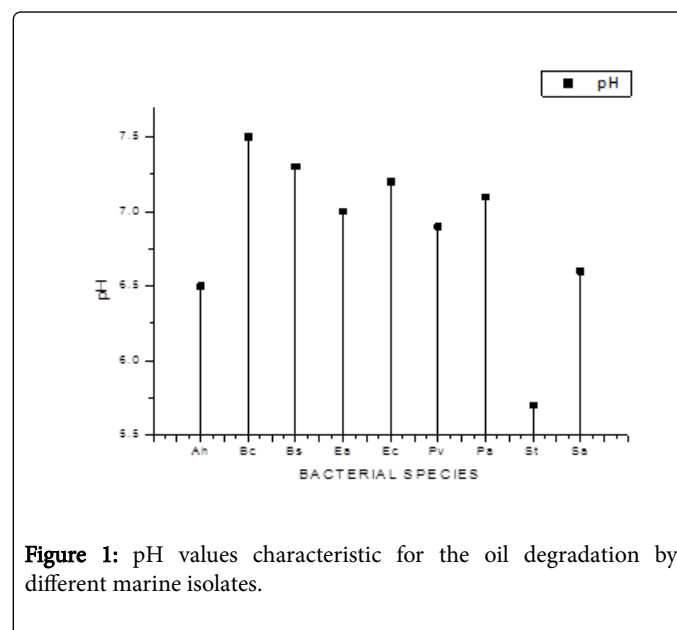


Figure 1: pH values characteristic for the oil degradation by different marine isolates.

Studies by Verstraete et al. [15] revealed that increase in pH fro 4.5 to 7.5 increased the bacterial capacity for biodegradation. However, the degradation capacity of the bacteria gradually decreased with increased pH above 9 of pH scale [16,17].

Degradation of oil

Study of the degradation of oil with isolated bacteria in different days were tabulated in Table 2. In our study degradation of oil began from 5th day onwards and maximum degradation by bacteria was observed on 11th day of our study compared to other days. The colony

forming units were observed more on 11th day and varied from 104 to 109 for different bacterial species. Affandi et al. determined that 10⁸ CFU ml⁻¹ was efficient for the degradation of oil and grease effluent from waste water from industries. In our study CFU values were in that range which resulted in the degradation of oil [18-20].

S.No	Name of the bacterial species	Percentage degradation bacteria (%)	of of	Amount of oil before inoculation of bacteria [ml]	Degradation of oil in days [ml]							
					3rd	5th	7th	9th	11th	13th	15th	17th
1.	<i>Aeromonas hydrophilia</i>	24		5.0	5.0	4.7	4.5	4.1	3.8	3.8	3.8	3.8
2.	<i>Bacillus cereus</i>	62		5.0	5.0	4.4	3.3	2.5	1.9	1.9	1.9	1.9
3.	<i>Bacillus subtilis</i>	52		5.0	5.0	4.6	3.7	3.0	2.4	2.4	2.4	2.4
4.	<i>Enterobacter aerogenes</i>	40		5.0	5.0	4.8	4.1	3.8	3.0	3.0	3.0	3.0
5.	<i>Escherichia coli</i>	46		5.0	5.0	4.7	3.9	3.1	2.7	2.7	2.7	2.7
6.	<i>Proteus vulgaris</i>	38		5.0	5.0	4.8	3.9	3.4	3.1	3.1	3.1	3.1
7.	<i>Pseudomonas auregenosis</i>	54		5.0	5.0	4.5	3.7	3.0	2.3	2.3	2.3	2.3
8.	<i>Salmonella typhimurium</i>	32		5.0	5.0	4.7	4.2	3.7	3.4	3.4	3.4	3.4
9.	<i>Staphylococcus aureus</i>	26		5.0	5.0	4.9	4.3	4.0	3.7	3.7	3.7	3.7

Table 2: Showing the degradation of oil with time period.

Conclusion

The present study determined the potential application of bacteria on the removal of oils in a cost effective and eco-friendly manner. The skill of native bacterial isolated from marine environment in the breakdown of oils would be advantageous and reduces the usage of normally expensive complex and synthetic growth medium. In our study it was identified that *Bacillus cereus* showed 62% of degradation of crude oil compared to other isolated strains. So, *Bacillus cereus* can be considered as potential oil degrader. Therefore, more studies are required to carry out in this line for better results of using native bacteria as an efficient tool for biodegradation of oil and related products.

References

- Leahy JG, Colwell RR (1990) Microbial degradation of hydrocarbons in the environment. *Microbiol Rev* 54: 305-315.
- Matsumiya Y, Wakita D, Kimura A, Sanpa S, Kubo M (2007) Isolation and characterization of a lipid-degrading bacterium and its application to lipid containing wastewater treatment. *J Biosci Bioeng* 103: 325-330.
- Nocentini M, Pinelli D, Fava F (2000) Bioremediation of a marine contaminated by hydrocarbon mixtures: the residual concentration problem. *Chemosphere* 41: 115-1123.
- Sakalle K, Rajkumar S (2009) Isolation of crude oil degrading marine bacteria and assessment for biosurfactant production. *Int J Microbiol* 7: 21-35.
- Mehdi H (2008) Investigation of alkane biodegradation using the microtiter plate method and correlation between biofilm formation, biosurfactant production and crude oil biodegradation. *International Biodeterioration and Biodegradation* 62: 170-178.
- Ramesh MA, Somashekar P (2014) Bioremediation of oil spill an invasion by bacteria to a safe environment. *Indian Journal of Applied Sciences* 4: 28-30.
- Gold T (1985) The origin of natural gas and petroleum, and the prognosis for future supplies. *Annu Rev Energy* 10: 53-77.
- Panda SK, Kar RN, Panda CR (2013) Isolation and identification of petroleum hydrocarbons degrading microorganisms from oil contaminated environment. *Int J Env Sc* 3: 1314-1321.
- Lidderdale T (1993) Demand, supply, and price outlook for low-sulfur diesel fuel. In *Energy Information Administration. Short Term Energy Outlook Annual Supplement DOE-E&0202*.
- Venkateswaran K, Iwabuchi T, Mastui Y, Toki H, Hamada E, et al. (1991) Distribution and biodegradation potential of oil-degrading bacteria in North Eastern Japanese coastal waters. *FEMS Microbiology Ecology* 1: 3-131.
- Prince RC, Clark JR, Lindstrom JE (1990) Bioremediation monitoring program of Alaskan oil spill. A publication of Exxon Research and Engineering, Annandale, Virginia, USA.
- APHA (2005) Standard methods for the examination of water and wastewater, 17th edn, American Public Health Association. Washington, USA
- Ahmad WA, Zakaria ZA, Zakaria Z, Surif S (2009) Hexavalent chromium reduction at different growth phases of *Acinetobacter haemolyticus*. *Environ Eng Sci* 26: 1275-1278.
- Nwuche CO, Ogbonna JC (2011) Isolation of lipase producing fungi from palm oil mill effluent (POME) dump sites at Nsukka. *Braz Arch Biol Technol* 54: 113-116.
- Verstraete W, Vanlooche R, DeBorger R, Verlinde A (1976) Modelling of the Breakdown and the Mobilization of Hydrocarbons in Unsaturated Soil Layers. *Applied Science Publishers, London, England* pp: 99-112.
- Cammarota MC, Teixeira GA, Freire DMG (2001) Enzymatic pre-hydrolysis and anaerobic degradation of wastewaters with high fat contents. *Biotechnology Letters* 23: 1591-1595.
- El-Bestawy E, El-Masry MH, El-Adi NE (2005) The potentiality of free Gram-negative bacteria for removing oil and grease from contaminating industrial effluents. *World Journal of Microbiology and Biotechnology* 21: 15-22.

18. Islam MS, Saiful M, Hossain M, Sikder M, Morshed M, et al. (2013) Acute toxicity of the mixtures of grease and engine wash oil on fish, pangasius sutch, under laboratory condition. Int J Lifesciences Biotechnology and Pharmacology Research 2: 306-317.
19. Singh A, Kumar V, Srivastava JN (2013) Assessment of bioremediation of oil and penol contents in refinery waste water via bacterial consortium. Journal of Petroleum and Environmental Biotechnology 4: 145
20. Vinothini C, Sudhakar S, Ravikumar R (2015) Biodegradation of petroleum and crude oil by *Pseudomonas putida* and *Bacillus cereus*. International Journal of Current Microbiology and Applied Sciences 4: 318-329.