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Microbial Community Dynamics Nutrient Recycling By Xenobiotics Degradation

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Abstract

The rate of glacier retreat has increased dramatically as a result of climate change. The ice has melted, exposing bare spots that can become new ecosystem development sites [1]. Here, primary microbial succession starts to build up organic material in preparation for plant colonisation. The Chhota Shigri glacier is one such glacier that has receded significantly and offers a perfect location for investigations of microbial succession [2]. Here, we looked at how the microbial communities and their functional characteristics changed as the glacier forefield transitioned from an exposed glacier snout to a fully-developed forested foreland [3]. Sequencing methods for metagenomes include amplicon sequencing [4]. Patescibacteria, Gemmatimonadota, Proteobacteria, Bacteroidota, and other microbial phyla were prevalent in the forefield locations closer to the glacier snout. These organisms have the capacity to cycle carbon and sulphur: Chloroflexi, Cyanobacteria, Verrucomicrobiota, and Myxococcota [5]. The heterotrophic taxa Actinobacteria and Acidobacteriota, which aid in the recycling of organic material, were prevalent at the places further from the glacier snout [6]. In comparison to locations farther from the glacier terminal, those closer to it had a greater variety and richness of microorganisms [7]. The whole-genome metagenome investigation also indicated the predominance of genes related to N, C, and cycle [8]. The local soil temperature was the main factor affecting the quantity and diversity of the microorganisms, followed by pH and element concentration. The bacteria and genes responsible for the breakdown of the xenobiotic chemicals Aminobenzoate, Benzoate, and Caprolactam have also been found in the soils of the forefield [9]. This study highlighted microbial successional gradients, which are caused by local environmental conditions [10].

Keywords: Microbial Succession; Functional Potential; Amplicon Sequencing; Whole-Genome Metagenome Sequencing

Introduction

Alpine glaciers react quickly to shifting weather patterns. Globally, rapid glacier retreat in alpine regions is one of the principal effects of rising global temperatures [11]. The exposed barren terrain created by the retreat of the glaciers provides a new home for a variety of microbial species. These microbes may be regarded as essential to the englacial ecosystem's operation since they are crucial to Furthermore, glaciers are not dead ecosystems; they contain a wide variety of microorganisms [12]. Upon glacial retreat, these microbial communities occupy the newly deglaciated region of the glacier and are beneficial in the main microbial succession and nutrient buildup. Studying alpine glacier forefields enables an understanding of the preliminary soil formation process along with primary microbial and plant succession processes along a defined chronosequence [13]. This is because reports from other alpine and subpolar glacier forefields have shown that primary microbial succession in the deglaciated site and the associated biogeochemical processes facilitate the colonisation of plants [14]. Along with aiding in the colonisation process, the deglaciated barren patches along a chronosequence aid in understanding how environmental change affects the early stages of succession [15]. Glacier forefields offer an intriguing setting for examining how microbial community distribution changes over time. Little research has been done on retreating glaciers and plant succession; data on microbial populations, however, since the main colonisers of the forefields of glaciers are still developing. The relationship between for exploration, glacier forefields offer a fascinating setting. Dispersion of the microbial community varying throughout time. There have been intermittent studies on retreating glaciers and plant succession, but the literature on microbial communities as the main colonisers of glacial forefields is still in its infancy. The microbial community's composition is determined by the soil's structure, and the microbial activities of

the community affect the soil's properties during the early stages of succession when bacteria aid in the mobilisation of nutrients through carbon and nitrogen fixing and rock weathering.

Discussion

Development, aiding the emergence and expansion of other organisms. Organisms to comprehend how and what exists in glacier forefield locations, it is critical to research the microbial composition and their functional features. Due to the lack of studies from various glaciers, it is still unclear how these local abiotic factors affect the primary succession of microbes at the glacier forefield. The current research was conducted to examine how local glacier environmental parameters affect the primary succession and the role of the microbes in nutrient recycling at the less-studied Western Himalayan glacier forefields. Since the studied glacier has retreated significantly, the deglaciated forefield areas will provide an ideal site for studying the microbial successional gradients. Additionally, the role of microorganisms in carbon and nitrogen cycling can be studied from such sites devoid of ice. Chhota Shigri Glacier, chosen as a benchmark glacier in the Western Himalaya for regional climate change study, has accelerated retreat. In the month of October, following the monsoon season, a total of twenty-four soil samples were taken from eight locations along the glacier forefield,

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commencing from close to the glacier snout to the forefield sites where vegetation was visible. The distance between each sample site is indicated in and a map of the glacier's snout using Sentinel2 data is provided in. The three soil samples were taken at locations that were five centimetres apart. The portion of the forefield where the soil was located was chosen for sample since the majority of the area was covered with rocks. For sampling, the top layer of soil was removed, and the soil between 5 and 10 cm deep was collected, properly sieved to remove big gravels, and then kept at 4 °C in sterile containers. Prior to each sample, ethanol. The dirt under the top layer was gathered since the deep soil layer is less susceptible to changes in the environment. Total DNA was extracted from the twenty-four forefield soil samples using a Fastenal spin kit for soil MP biomedical, California, USA in accordance with manufacturer protocol. The soil temperature onsite was measured using a digital thermometer Mixtec Technologies India Private Limited at a depth of the pH and conductivity of the soil samples were checked using a pH and conductivity measurement device. The DNA that was isolated has good quality. The extracted DNA from the seven spots in triplicates was verified on an agarose gel, and quantification was carried out using Nano Drop One. HAM39 soil sample failed the QC, Thermos Scientific's Gene JET Gel Extraction Kit was used to purify the amplified products to eliminate non-specific amplified products, and the NEBNext Ultra DNA library preparation kit was then used to prepare the libraries New England Biolabs, UK. Using the Agilent 2200 TapeStation, the quality and amount of the DNA library were evaluated.

Conclusion

Agilent Technologies, USA, used the Illumina HiSeq 2500 platform for the sequencing. For the purpose of whole genome metagenome sequencing, the DNA collected from the initial four sites and the final four forefield sites was combined. The Native barcoding genomic DNA kit was used to barcode the DNA from the combined samples. Soil samples were taken in triplicate at locations that were 5 cm apart. Rocks covered the forefield area for the most part, The soil under 5 to 10 cm of topsoil was gathered, carefully sieved to remove big gravels, and kept at 4 in sterile zip-lock bags until further use. Prior to each sampling, 70% ethanol was used to sterilise the shovel and sieve that were utilised for the sample. The dirt under the top layer was gathered since the deep soil layer is less susceptible to changes in the environment. A digital thermometer from Mextech Technologies India Private Limited was used to assess the soil temperature there at a depth of Microbial alpha diversity indices Observed, Chao-1, Shannon, and Simpson were tested; they all showed a consistent pattern throughout the forefield locations. There was relatively increased bacterial richness at locations, according to the observed species index, which displays the total number of species in the sites. Across the glacier forefield locations, there were also variations in bacterial beta diversity. In contrast, the abundance of Patescibacteria, Gemmatimonadota, Proteobacteria, Bacteroidota, Chloroflexi, Cyanobacteria, Verrucomicrobiota, and Myxococcota has decreased across the forefield sites, suggesting that these phyla are more abundant at sites close to the glacier terminus glacier forefield. The abundance of the genera Actinobacteria and Acido The research discovered the existence of genes involved in the metabolism of lipids, nitrogen, sulphur, and nucleotides. Interestingly, the glacial forefield metagenome also contained genes involved in xenobiotic biodegradation. The glacial metagenome included the genes and proteins essential for C metabolism and cycling. In the soil samples distant, there were more genes/proteins involved in photosystems I and II photosynthesis. The genes essential for prokaryotic carbon fixation

pathways and methane metabolism were similarly common in both the metagenomic sample. 'The examined forefield soil metagenome contained genes involved in organic N metabolism and recycling in both samples. A complete list of the numerous genes and their enzymes involved in carbon metabolism is provided in. The dissimilatory nitrate reduction genes were widely distributed in both metagenomes. In contrast, the genesnirB were more common in the sample far from the glacier's terminal than in the one close to it. Moreover, both metagenomic samples contained the genes necessary for assimilatory nitrate reduction. In locations far from the terminal, genes involved in the denitrification process predominated.

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Conflict of Interest

None

References

- 1. Argumedo Delira R, Gómez Martínez MJ, Soto BJ (2019) [Gold Bioleaching](https://www.researchgate.net/publication/343807899_Microorganisms_and_Plants_in_the_Recovery_of_Metals_from_the_Printed_Circuit_Boards_of_Computers_and_Cell_Phones_A_Mini_Review) [from Printed Circuit Boards of Mobile Phones by Aspergillus niger in a Culture](https://www.researchgate.net/publication/343807899_Microorganisms_and_Plants_in_the_Recovery_of_Metals_from_the_Printed_Circuit_Boards_of_Computers_and_Cell_Phones_A_Mini_Review) [without Agitation and with Glucose as a Carbon Source.](https://www.researchgate.net/publication/343807899_Microorganisms_and_Plants_in_the_Recovery_of_Metals_from_the_Printed_Circuit_Boards_of_Computers_and_Cell_Phones_A_Mini_Review) Metals 9: 521.
- 2. Bindschedler, Bouquet, TQT V, Job D, Joseph, et al. (2017) [Fungal biorecovery](https://www.semanticscholar.org/paper/Fungal-Biorecovery-of-Gold-From-E-waste.-Bindschedler-Bouquet/99137daff794a7f51bc384f657c95fb4f876614f) [of gold from e-waste.](https://www.semanticscholar.org/paper/Fungal-Biorecovery-of-Gold-From-E-waste.-Bindschedler-Bouquet/99137daff794a7f51bc384f657c95fb4f876614f) Adv Appl Microbiol 99: 53-81.
- 3. Dave SR, Sodha AB, Tipre DR (2018) [Microbial technology for metal recovery](https://www.researchgate.net/publication/328176386_Microbial_technology_for_metal_recovery_from_e-waste_printed_circuit_boards) [from e-waste printed circuit boards](https://www.researchgate.net/publication/328176386_Microbial_technology_for_metal_recovery_from_e-waste_printed_circuit_boards). J Bacteriol Mycol Open Access 6: 241-247.
- 4. Debnath, Biswajit, Chowdhury, Ranjana Ghosh, Sadhan, et al. (2018). [Sustainability of metal recovery from E-waste.](https://www.researchgate.net/publication/328176386_Microbial_technology_for_metal_recovery_from_e-waste_printed_circuit_boards) Front Environ Sci 1044-1109.
- 5. Díaz Martínez ME, Argumedo Delira R, Sánchez Viveros G (2019) [Microbial](https://www.researchgate.net/publication/331294557_Microbial_Bioleaching_of_Ag_Au_and_Cu_from_Printed_Circuit_Boards_of_Mobile_Phones) [Bioleaching of Ag, Au and Cu from Printed Circuit Boards of Mobile Phones.](https://www.researchgate.net/publication/331294557_Microbial_Bioleaching_of_Ag_Au_and_Cu_from_Printed_Circuit_Boards_of_Mobile_Phones) Curr Microbiol 76: 536-544.
- 6. Forti V, Baldé CP, Kuehr R, Bel G, Monitor GEW, et al. (2020). [Quantities,](https://www.itu.int/en/ITU-D/Environment/Documents/Toolbox/GEM_2020_def.pdf) [flows and the circular economy potential](https://www.itu.int/en/ITU-D/Environment/Documents/Toolbox/GEM_2020_def.pdf). The Global E waste Monitor 13-15.
- 7. Li J, Xu T, Liu J, Wen J, Gong S, et al. (2021) [Bioleaching metals from waste](https://www.researchgate.net/publication/44661795_Thermodynamic_Analysis_of_Contamination_by_Alloying_Elements_in_Aluminum_Recycling) [electrical and electronic equipment \(WEEE\) by Aspergillus niger: a review.](https://www.researchgate.net/publication/44661795_Thermodynamic_Analysis_of_Contamination_by_Alloying_Elements_in_Aluminum_Recycling) Environ Sci Pollut Res 28: 44622-44637.
- 8. Madrigal Arias JE, Argumedo Delira R, Alarcón A, Mendoza López M, García Barradas, et al. (2015) [Bioleaching of gold, copper and nickel from waste](https://www.mdpi.com/2075-4701/9/5/521/htm) [cellular phone PCBs and computer goldfinger motherboards by two Aspergillus](https://www.mdpi.com/2075-4701/9/5/521/htm) [niger strains](https://www.mdpi.com/2075-4701/9/5/521/htm). Braz J Microbiol 46: 707-713.
- 9. Giovannoni SJ, Britschgi TB, Moyer CL, Field KG (1990) [Genetic diversity in](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8547001/) [Sargasso Sea bacterioplankton.](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8547001/) Nature 345: 60–63.
- 10. Parks DH, Chuvochina M, Waite DW, Rinke C, Skarshewski A, et al. (2018) [A standardized bacterial taxonomy based on genome phylogeny substantially](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8547001/) [revises the tree of life.](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8547001/) Nat Biotechnol 36: 996-1004.
- 11. Hug LA, Baker BJ, Anantharaman K, Brown CT, Probst AJ (2016) [A new view](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8547001/) [of the tree of life](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8547001/). Nat Microbiol 1: 16048.
- 12. Whitman WB, Oren A, Chuvochina M, da Costa MS, Garrity GM, et al. (2018) [Proposal of the suffix –ota to denote phyla. Addendum to 'Proposal to include](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8547001/) [the rank of phylum in the International Code of Nomenclature of Prokaryotes.](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8547001/) Int J Syst Evol Microbiol 68: 967-969.
- 13. Hugenholtz P, Tyson GW, Webb RI, Wagner AM, Blackall LL, et al. (2001) [Investigation of candidate division TM7, a recently recognized major lineage of](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC92593/) [the domain Bacteria, with no known pure-culture representatives.](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC92593/) Appl Environ Microbiol 67: 411-419.
- 14. Mummey DL, Stahl PD (2003) [Candidate division BD: phylogeny, distribution](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8547001/) [and abundance in soil ecosystems.](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8547001/) Syst Appl Microbiol 26: 228-235.
- 15. Zhang H, Sekiguchi Y, Hanada S, Hugenholtz P, Kim H, et al. (2003) [Gemmatimonas aurantiaca gen. nov., sp. nov., a Gram-negative, aerobic,](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8547001/) [polyphosphate-accumulating micro-organism, the first cultured representative](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8547001/) [of the new bacterial phylum Gemmatimonadetes phyl.](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8547001/) nov Int J Syst Evol Microbiol 53: 1155-1163.