

Changing of a Single Core Gene, *tssM*, of Type VI Secretion System of *Xanthomonas perforans* Influences Virulence, Epiphytic Survival, and Transmission during Pathogenesis on Tomato

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Editorial

In eukaryotes, protein phosphatases play a vital role in the control of different cellular activities. *Colletotrichum gloeosporioides*, an ascomycete, is the cause of anthracnose disease in a number of key crops and plants. *CgPPZ1*, a protein phosphate gene and a homolog of yeast *PPZ1*, was discovered in *C. gloeosporioides* in this investigation. *CgPpz1* was found to be critical for vegetative growth and asexual development, conidial germination, and plant infection after targeted gene deletion. *CgPpz1* was found to be located in the cytoplasm during cytological tests. Osmotic stresses, cell wall stressors, and oxidative stressors were all hypersensitive in the *Cgppz1* mutant. Our findings suggest that *CgPpz1* is involved in the fungal development and pathogenicity of *C. gloeosporioides*, as well as the numerous stress responses that occur [1].

Xanthomonas perforans is a seedborne hemibiotrophic pathogen that infects the tomato phyllosphere. While most investigations on the molecular basis of pathogenesis have focused on apoplastic expansion, variables relevant during asymptomatic colonisation in the early stages of disease development are little known. The *tssM* gene of the type VI secretion system cluster *i3** (*T6SS-i3**) plays an important role during initial asymptomatic epiphytic colonisation at various phases during the pathogen's life cycle, according to this study. When dip inoculating 4- to 5-week-old tomato plants, a mutation in a key gene, *tssM* of *T6SS-i3**, conferred stronger pathogen aggressiveness, as seen by higher overall disease severity, higher in planta growth, and shorter latent infection period compared to the wild-type [2]. The role of *tssM* in aggressiveness was demonstrated during vertical transmission from seed to seedling, with wild-type seedlings demonstrating lower disease severity and in planta populations on seedlings than mutants. When examined in an experimental system that mimicked transplant house high-humidity conditions, the presence of functioning *TssM* resulted in enhanced epiphytic fitness as well as greater pathogen dissemination potential. We discovered that one mechanism by which *TssM* promotes epiphytic fitness is through increased osmotolerance. These findings suggest that a functional *TssM* plays a significant role in the pathogen's ability to gain an ecological advantage. *TssM* prolongs the hemibiotrophic pathogen's relationship with the host, reducing disease severity while allowing successful diffusion [3].

The pears (*Pyrus* spp.) are one of China's most popular fruits, but their supply is under threat from deadly illnesses. In this paper, we describe two cases of stem canker and twig dieback disease on pear plants in Guangxi and Jiangsu provinces, which resulted in the death of pear seedlings (about 10% of total plants). The fungus *Neofusicoccum parvum* was identified as the disease's causal agent in these two locations using a combination of morphological and molecular diagnostics, as well as a pathogenicity test [4]. The isolates were classified into two clades: the CY-2 isolate and the other four isolates, ZL-4, BM-9, BM-10, and BM-12, may have split into two *N. parvum* groupings. For further examination, two representative isolates (CY-2 and ZL-4) were chosen.

The best temperature for in vitro infection on pear trees of these two isolates was around 25°C, according to our findings [5]. In vitro, both CY-2 and ZL-4 could infect many sand pear cultivars as well as other horticultural plants, however CY-2 was more virulent on certain pear varieties, including Nanyue, Lvyun, Qiushui, and Ningmenghuang. Additionally, the efficacy of fungicides against these two isolates was assessed, with carbendazim and flusilazole proving to be the most efficient fungicides in preventing the growth of these fungal infections. These findings, taken together, characterise the *N. parvum* species and offer viable solutions for the disease's future management [6].

Physiological saline solution was used to wash the germs away from the surface of the leaves or cones. The extract was used in different dilutions of a bacterial, fungal, and yeast culture medium. Sequential sprouting on agar plates yielded pure microorganism cultures. The Biolog microbiological ID system was used to identify all of the bacteria species (Hayward, CA) [7]. There are now 2,700 bacterium types in the database. Microscopic observations and the physical appearance of the colonies on the dishes were used to identify the fungi on a generic level. The fact that they have been identified is simply a hint. *Bacillus* species (*megatherium*, *idriensis*, *indicus*, *endophyticus*, *licheniformis*) and *Pantotea* species (*aglomerans*, *dispersa*) were the most commonly identified in the bacterial population in both years [8]. Fungi have a lot more variation, which is probably due to the diametrically opposite weather conditions in 2015 and 2016. In both years, just two fungi from the genera *Penicillium* and *Rhizopus* were found out of a total of eight. Yeasts were only found on hop leaves on rare occasions. Bacteria were discovered in both dried hops and hop pellets. This proves that post-harvest hops processing, kiln drying, and pelletization are not sterilising methods [9].

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Conflict of Interest

No potential conflicts of interest relevant to this article were reported.

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