

commentary

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# Relink Stk11/Lkb1 in Stromal Cells to Peutz-Jeghers Syndrome

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## Description

Lkb1 (Liver Kinase B1, encoded by Stk11) is a multifaceted serine/ threonine kinase recognized as a tumor suppressor, regulating cell metabolism, cell polarity, cell fate, and cell survival [1-6]. Previous studies revealed that global knockout of Stk11 in mice leads to embryonic lethality, indicating the pivotal role of Lkb1 in embryonic development [7]. Intriguingly, it is widely accepted that germline mutations in Stk11 are strongly associated with Peutz-Jeghers Syndrome (PJS) in humans, an inherited disease characterized by Gastrointestinal (GI) hamartomatous polyposis and increased risks for multiple types of cancers [8]. Subsequently, the exciting findings to manifest PJS-like intestinal polyposis in mouse models with heterozygous deletion of Stk11, have fueled the researchers worldwide to pursue the specific role of Lkb1 in the pathogenesis of PJS [9-11].

More and more conditional *Lkb1* deletion mouse models have been developed to open new avenues for further investigation of the cell types and signaling pathways involved in polyp formation. Surprisingly, deletion of *Lkb1* in intestinal epithelial cells results in increased susceptibility to colitis and suppression for microbial population, but shows no evidence of GI polyps in mice, even those 52 weeks of age [12,13]. These lines of evidence indicate the potential roles of nonepithelial *Stk11/Lkb1* in the development of PJS-associated GI polyps. To this end, multiple conditional knockout mouse models of *Lkb1* in mesenchymal (stromal) cells have been generated, including Tagln-Cre (Smooth Muscle Cell (SMC)-specific), Fsp1-Cre (Fibroblast -specific), Twist2-Cre and Gli1-Cre (mesenchymal progenitor cell-specific) Nkx3.2-Cre (pan-mesenchymal cell specific), which suggest that *Lkb1* mutation in certain murine stromal cells could drive PJS-like GI polyposis N [14-16].

Our recent study further confirmed the critical role of mesenchymal Stk11/Lkb1 in the pathogenesis of gastrointestinal polyposis [17]. We generated tamoxifen-inducible  $Lkb1^{flox/+};Myh11-Cre/ERT2$  (Lkb1 Het) and  $Lkb1^{flox/flox};Myh11-Cre/ERT2$  (Lkb1 KO) mice. We found that heterozygous rather than homozygous Lkb1 deletion in murine mature SMCs is sufficient for the manifestation of PJS-like polyps, which is inconsistent with previous finding observed in mice with SMC-targeted inactivation of Stk11 by Tagln-Cre [14]. PJS-like polyps, characterized by an arborizing smooth muscle core, abundant ECM deposition and augmented immune cell infiltration, were observed in Lkb1 Het mice from 9 months post-tamoxifen treatment, in contrast to none developed in Lkb1 KO mice till their death. Furthermore,  $Lkb1^{flox/flox};Pdgfra-Cre/ERT2$  mice, another mesen- chymal Stk11/Lkb1 deletion model, also simulated historically similar polyps to those in Lkb1 Het GI, as early as 2-3 months after tamoxifen treatment. Results supported the notion

that Myh11<sup>+</sup> or Pdgfrα<sup>+</sup> mesenchymal cells may serve as an important cellular origin for PJS-like polyps.

To provide novel insights into the comprehensive cellular components and the underlying molecular mechanisms of the Lkb1associated polyps, we performed a single-cell transcriptome atlas of Lkb1-associated polyps for the first time in Lkb1<sup>flox/+</sup>; Myh11-Cre/ERT2 mice. Clustering analysis revealed that there are polyposis-specific cell clusters and a higher portion of mesenchymal cells within Lkb1 Het duodenum polyps, compared with normal GI tissues. As the largest cell population in duodenum, the epithelial cells from Lkb1 Het polyp exhibited aberrant stem cell-like characteristics at an impaired differentiation state, along with an increment in expression of stem cell markers such as Cd44 as previously clarified but a decrement in mature enterocyte markers [15,18]. Of note, the up regulation of genes encoding secretory proteins of the gastric mucus barrier in those abnormal stem cell-like epithelial cells displayed the functional switch into a more secretary phenotype consistent with previous findings, which necessitates further research into the biological significance of Lkb1 in GI homeostasis.

Interestingly, coupled with the reported Spp1-Cd44 axis promoting tumor progression and metastases we found that intercellular communication networks (Spp1-Cd44 or Spp1-Itga8/Itgb1) among the epithelial, mesenchymal/stromal, and immune cells contribute to polyposis process [19,20]. Besides, special focus should be given to the abundant immune cell infiltration in *Lkb1*-related polyps in our study and other studies. Previous study demonstrated that *Lkb1* deficiency in T cells is sufficient to promote the development of gastrointestinal polyps [13]. However, the underlying mechanism of deregulated inflammatory responses caused by *Stk11/Lkb1* inactivation in stromal cells and immune cells is awaited to further identified.

In general, it is the first time to conduct a single-cell transcriptome atlas of *Lkb1*-associated polyps, trying to elucidate the pathological microenvironment changes, variations in cellular constitutions and functionalities and possible signaling pathways in cell-cell interactions. Key questions remain to be answered about how mesenchymal *Lkb1* regulates epithelial cell fate/state in *Lkb1*-associated polyps. Further research is warranted in the aim of yielding clinical benefits for patients with PJS.

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#### References

- 1. Shaw RJ, Lamia KA, Vasquez D, Koo SH, Bardeesy N, et al. (2005) The kinase *LKB1* mediates glucose homeostasis in liver and therapeutic 12. effects of metformin. Science 310:1642-1646.
- Fu A, Ng AC, Depatie C, Wijesekara N, He Y, et al. (2009) Loss of *Lkb1* in adult beta cells increases beta cell mass and enhances glucose 13. tolerance in mice. Cell Metab 10:285-295.
- 3. Yang K, Blanco DB, Neale G, Vogel P, Avila J, et al. (2017) Homeostatic control of metabolic and functional fitness of Treg cells by *LKB1* signalling. Nature 548:602-606.
- Shorning BY, Zabkiewicz J, McCarthy A, Pearson HB, Winton DJ, et al. (2009) Lkb1 deficiency alters goblet and paneth cell differentiation in the 15. small intestine. PLoS One 4:e4264.
- 5. Wu D, Luo Y, Guo W, Niu Q, Xue T, et al. (2017) Lkb1 maintains Treg cell lineage identity. Nat Commun 8:15876.
- Cai Z, Satyanarayana G, Song P, Zhao F, You S, et al. (2024) Regulation of Ptbp1-controlled alternative splicing of pyruvate kinase muscle by Liver kinase b1 governs vascular smooth muscle cell plasticity *in vivo*. 17. Cardiovasc Res.
- Ylikorkala A, Rossi DJ, Korsisaari N, Luukko K, Alitalo K, et al. (2001) Vascular abnormalities and deregulation of VEGF in *Lkb1*-deficient mice. Science 293:1323-1326.
- Lim W, Olschwang S, Keller JJ, Westerman AM, Menko FH, et al. (2004) Relative frequency and morphology of cancers in *STK11 mutation carriers*. Gastroenterology;126:1788-1794.
- 9. Bardeesy N, Sinha M, Hezel AF, Signoretti S, Hathaway NA, et al. (2002) Loss of the *Lkb1* tumour suppressor provokes intestinal polyposis but resistance to transformation. Nature 419:162-167.
- 10. Miyoshi H, Nakau M, Ishikawa TO, Seldin MF, Oshima M, et al. (2002) Gastrointestinal hamartomatous polyposis in *Lkb1* heterozygous knockout mice. Cancer Res 62:2261-2266.

- 11. Jishage K, Nezu J, Kawase Y, Iwata T, Watanabe M, et al. (2002) Role of Lkb1, the causative gene of Peutz-Jegher's syndrome, in embryogenesis and polyposis. Proc Natl Acad Sci 99:8903-8908.
- Liu Xn, Lu J, Liu Z, Zhao J, Sun H, et al. (2018) Intestinal epithelial cellderived *LKB1* suppresses colitogenic microbiota. J Immunol 200:1889-1900.
- Poffenberger MC, Metcalfe-Roach A, Aguilar E, Chen J, Hsu BE, et al. (2018) *LKB1* deficiency in T cells promotes the development of gastrointestinal polyposis. Science 361:406-411.
- Katajisto P, Vaahtomeri K, Ekman N, Ventelä E, Ristimäki A, et al. (2008) *LKB1* signaling in mesenchymal cells required for suppression of gastrointestinal polyposis. Nat Genet 40:455-459.
- Ollila S, Domènech-Moreno E, Laajanen K, Wong IPL, Tripathi S, et al. (2017) Stromal *Lkb1* deficiency leads to gastrointestinal tumorigenesis involving the IL-11-JAK/STAT3 pathway. J Clin Invest128:402-414.
- Cotton JL, Dang K, Hu L, Sun Y, Singh A, et al. (2022) PTEN and LKB1 are differentially required in *GLI1*-expressing mesenchymal cells to suppress gastrointestinal polyposis. Cell Rep 40:111125.
- Cai Z, Jiang Y, Tong H, Liang M, Huang Y, et al. (2024) Cellular and molecular characteristics of stromal *Lkb1* deficiency-induced gastrointestinal polyposis based on single-cell RNA sequencing. J Pathol 263:47-60.
- Lai C, Robinson J, Clark S, Stamp G, Poulsom R, et al. (2011) Elevation of WNT5A expression in polyp formation in Lkb1 ± mice and Peutz-Jeghers syndrome. J Pathol 223:584-592.
- Nallasamy P, Nimmakayala RK, Karmakar S, Leon F, Seshacharyulu P, et al. (2021) Pancreatic tumor microenvironment factor promotes cancer stemness via SPP1-CD44 Axis. Gastroenterology 161:1998-2013.
- 20. Rao G, Wang H, Li B, Huang L, Xue D, et al. (2013) Reciprocal interactions between tumor-associated macrophages and CD44-positive cancer cells *via* osteopontin/CD44 promote tumorigenicity in colorectal cancer. Clin Cancer Res 19:785-97.