

Microorganisms Metabolism during Bioremediation of Oil Contaminated Soils

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Abstract

The development of some industrial processes, such as extraction, refining and oil transportation has been the cause of environmental degradation caused by oil spills and their extraction processes involved. In order to reduce these contaminants, among others, physicochemical technologies have been applied, which haven't solved the contamination problem because in many of these processes, new toxic compounds are generated. Technologies based in the use of microorganisms, plants and other biological species are called bioremediation. This is an alternative to the physical and chemical processes of treatment that, although slower in response, are more sustainable. Therefore, in this review, bioremediation technologies to treat oil contaminated soils are analyzed as a convenient alternative for the restoration of impacted soils, against progressive deterioration of environmental quality. Additionally, the microorganisms as well as the required conditions for which bioremediation is considered a sustainable process to undertake will also be presented.

Keywords: Bioremediation; Microorganisms; Hydrocarbons

Introduction

Soil is a natural nonrenewable resource; it is the most easily affected space of the oil industry (exploration, drilling and exploitation) because of the residue the industry generates. In countries where oil activity is fundamental, the ground and subsurface is highly polluted [1]. This contamination can be carried to the aquifers; this is why soil rehabilitation is a priority.

There are different physicochemical techniques to control and mitigate hydrocarbon contamination. Among the methods used to restore contaminated ground there is incineration, thermal desorption, and ground confinement to locations destined for its dispose [2]. Although considered effective, incineration and desorption strategies prompt harsh damage to the soil; the treatment causes losses in its nutritional value. These methods also generate carbon dioxide, carbon monoxide, sulfur dioxide and nitrogen oxides, which are gasses with greenhouse effect [2]. The confinement method consists in removing residue and/or contaminated soil and releasing it to a localized area or in covering it with an impermeable coating, that allows the permanence of hydrocarbon in the subsoil, which lixivates to the shallow groundwater reserves or spread on the surface.

These methods are expensive, cost time and are not a real solution to the problem on the long run [3]. In the past years, the use of biological technologies has demonstrated to be efficient and adequate, and cause less impact to the location where the problem is found. These techniques are denominated bioremediation, which is the technology that uses microorganisms' natural metabolic potential to transform contaminants into simple compounds, that are slightly or non-pollutant. Bioremediation uses environmentally sustainable methods [4]. Bioremediation does neither alter nor modify the food chain in the treated ecosystems: it respects the flora and fauna of the contaminated site. This is a biotechnological practice that has gained great importance worldwide, because it is friendly with the environment and very low in cost [5]. The following example demonstrates the value in dollars for each one of the treatments applied in one ton of contaminated soil (value in dollars): deposit or confinement: \$140-200, incineration: \$140-150, stabilization: between \$100 y 200 and bioremediation:

between \$15 y 170 [6]. The application of bioremediation techniques reduce or eliminate the costs for soil transportation because the ground can be treated directly in the contaminated area. This is a feasible technique that can be applied in areas that are difficult to access, where, once the site has been cleansed, it may be once more used as landfill for industrial areas, if completely restored, it may be used for housing developments and, in some cases, even for agricultural purposes.

Toxicity in contaminated ground by hydrocarbon has been proven by toxicity bioassays, which can provide information about the characterization process and the ecological risks in the contaminated site. Toxicity in petroleum hydrocarbons is strongly correlated with a low boiling point and especially with those that are within the range of C10-C19. Some metabolic compounds such as aldehydes, aromatic ketones, carboxylic acids, fatty acids and esters, also contribute to the contamination of the soil. There is proof that allows the evaluation of the toxicity in soil, such as Microtox, the use of earthworms and seed germination trials. The time of exposition of the ground to it may favor the decrease of toxicity in the ground by means of the humic acid activity in the soil. However, the change in the toxicity under different conditions and time exposition is still unknown because of the intervening variables [7].

Hydrocarbon biodegradation

The process of digestion, assimilation and hydrocarbon metabolism

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carried out by bacteria, fungi, protozoa and other organisms (Figure 1). The process of biodegradation includes the following reactions: oxidation-reduction, adsorption processes and ion exchange, as well as chelation reactions that result in metal buildup. Microbial biodegradability allows the transformation of hydrocarbons of complex structure into a more simple chemical structure [8]. When the transformation is simple, it is called "primary", when complete, "mineralization", in the latter, hydrocarbon is decomposed into inorganic compounds and/or cellular constituents [9,10]. The acceleration in hydrocarbon biodegradation may be carried out by native microorganisms or bio-augmentation, with appropriate nutritional and environmental conditions [11].

Hydrocarbon biodegradative microorganisms

Bacteria: Among the most studied bacteria in this process are *Pseudomonas*, *Alcaligenes*, *Sphingomonas*, *Rhodococcus* and *Mycobacterium*, these degrade aliphatic and aromatic hydrocarbons, using them as the only source of carbon and energy. Aerobic degradation is carried out by dioxygenases enzymes that use oxygen during the degradation process (Figure 2). Microorganisms first oxidize the carbon molecule through a complex multi-enzymatic (oxygenase type enzymes) that incorporate an oxygen molecule. This is how a hydrocarbon with an alcohol group molecule (a more reactive one) is obtained. Other enzymes oxidize the alcohol group to an aldehyde group and finally to a carboxylic acid. Because of this, a molecule, similar to a fatty acid, is obtained and it may be degraded to an acetyl-CoA by beta oxidation. This process of oxidation can occur in non-terminal carbons, giving place to two fatty acids that are processed by beta oxidation [12].

Biodegradation can also be done in the absence of oxygen, as it happens in the deepest hydrocarbon sediments or reservoirs from which anaerobic bacteria has been isolated [13,14]. Bacteria use nitrates, sulfates and iron as electron acceptors for their metabolism [15]. Example of it is the *Archaeas* bacteria that oxidize methane in association with sulfate reducing bacteria [16]. Other anaerobic bacteria that have been reported with hydrocarbon biodegradable capacity are *Dechloromonas aromatica* and some species from the genus *Azoarcus*, *Thauera* [17,18].

Fungi: These microorganisms are known for their diversity and for their capacity to degrade natural, persistent and complex material [19]. The mycelium contains insoluble substances. With them, it begins the process of biodegradation, allowing for subsequent entrance or succession of other microorganisms and facilitating compound decomposition [20]. Fungi can extend by means of their hyphae and grow in stress conditions that bacteria cannot resist due to low disposition to nutrients, low pH and low humidity [1]. In hydrocarbon bioremediation, the fungi of white rot (lignin fungi) have been mostly studied [21,22]. Among these species there is *Tyromyces palustris*, *Gloeophyllum trabeum*, *Trametes versicolor*, which oxidize hydrocarbons by the action of the laccase enzyme [23]; *Phanerochaete chrysosporium*, oxidizes the anthracene and the pyrene through lignin peroxidase enzymes (LiP) and manganese peroxidase (MnP) [24,25], *Stropharia coronilla* oxidizes benzopyrene through MnP [26] and *Pleurotus ostreatus* which decomposes the benzantracene, chrysene, benzofluoranthene, benzopyrene, dibenzanthracene and benzoperylene through the laccase and the MnP [27].

Filamentous fungi, native to the ground [21] can also be decomposed; however, in many cases they must first be adapted by developing them on solid supports (sugar cane bagasse, sorghum or

other agricultural residue) [28,29], with small contaminant quantities in order to place them later on the contaminated site. This process of bioaugmentation may also be applied to the lignin fungi [30].

Yeast: Yeasts may also be used in bioremediation of hydrocarbons, among which there are *Debaryomyces*, *Saccharomyces*, *Yarrowia*, *Pichia* and some species of *Candida* [31,32]. Yeast *Candida viswanathii* has demonstrated a high percentage of removal of phenanthrene (77.21-89.76 %), as this it is a low- molecular weight hydrocarbon, it also demonstrated high efficiency of removal (55.53-60.77 %) of benzopyrene, which is a high-molecular weight hydrocarbon [33]. The isolated strain *Pichia anomala* 2.2540, from a contaminated ground, demonstrated high removal at 96 hours, in cultures with naphthalene, dibenzothiophene, phenanthrene and chrysene, both in individual as in combined forms [34]. Yeasts have the capacity of adaptability, such is the example of the *Candida digboiensis*, strain, isolated from the contaminated ground by hydrocarbons and acidic pH [35].

Algae: Walker et al. in 1975 [36] were the first to report the isolation of algae *Prototheca zopfii*, which decomposes petroleum and other

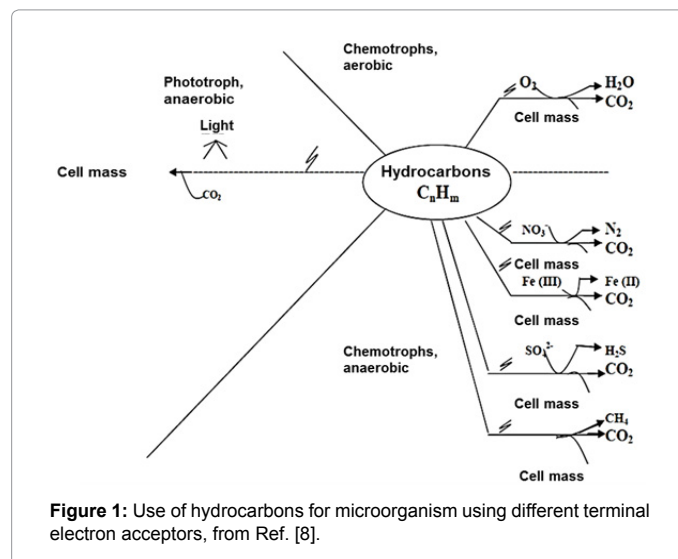


Figure 1: Use of hydrocarbons for microorganism using different terminal electron acceptors, from Ref. [8].

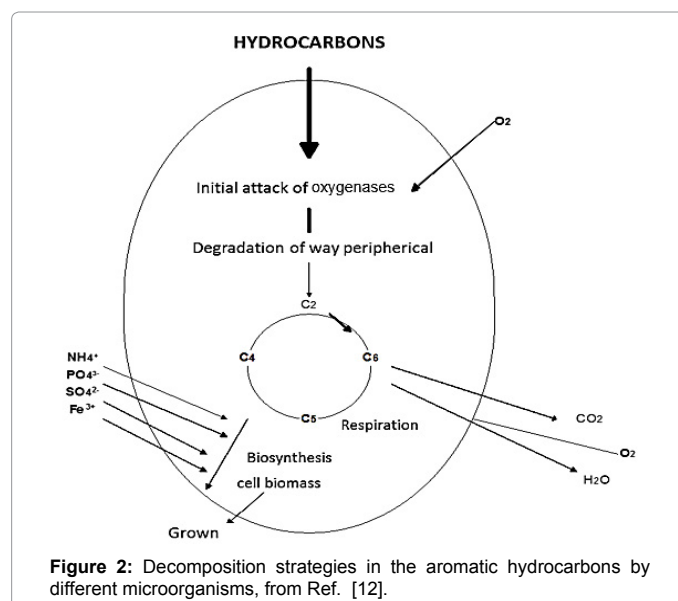


Figure 2: Decomposition strategies in the aromatic hydrocarbons by different microorganisms, from Ref. [12].

hydrocarbons. Marine algae (cyanobacteria, green algae and diatoms) are capable of metabolizing naphthalene into a series of metabolites [37]. The metabolic route of decomposition of hydrocarbons of the genus *Oscillatoria* and *Agmenellum spp* is similar to that of the fungi. In addition, algae possess an important potential in the consortia with bacteria, for example: *Pseudomonas migulae* and *Sphingomonas yanoikuyae* that decompose phenanthrene. Microalgae *S. costatum* and *Nitzschia sp.* can accumulate and decompose two polycyclic hydrocarbons simultaneously [38].

Protozoa: The contribution of protozoa in bioremediation of hydrocarbons is considered controversial since it does not have the capacity to use them directly as an energy source: protozoa are considered to have a negative effect, due to the fact that they feed off the bacterium that contributes to decomposing hydrocarbons. However, other investigations consider that protozoa have a positive effect in the bioremediation systems, because they contribute to keep porosity in the contaminated ground by regulating bacterial count [39].

Microbial consortia: In nature, microorganisms are generally found in consortia that interact synergistically, allowing a greater removal of pollutants in the ground [40,41]. When exposed to contaminated areas during long periods of time, the microbial community present in the ground adapts to the pollutants [42]. This benefits the process of bioremediation. Microbial consortia have proven to render satisfactory results in reducing contaminants in ground that was previously isolated [43]. Nevertheless, there are consortia that have been tested in extreme conditions because of its adaptability. Such is the case of the consortia formed by *Acinetobacter sp.*, *Pseudomonas sp.*, *Ralstonia sp.* and *Microbacterium sp.* which decompose aromatic hydrocarbons of high-molecular weight at low temperatures [44]. The enzyme that is activated for the biodegradation of the hydrocarbon depends on the microorganism and the type of hydrocarbon (Table 1) [45].

Microbial metabolism for the decomposition of hydrocarbons: In the processes of bioremediation there are interactions between the enzymes that degrade hydrocarbons, which modify their structure as well as their toxicity, in aerobic or anaerobic conditions. This depends on the systems conditions and the microorganisms that develop in it [61].

Aerobic route: The aerobic degradation is carried out within the cytoplasmic membrane or on the cellular surface. The product of oxidation is transported inside the cell [62], where the microorganisms oxidize the carbon molecule through a multi-enzyme complex (oxygenases enzymes) that incorporates an oxygen molecule [59]. This results in a hydroxylated hydrocarbon (alcohol), a more reactive molecule. Subsequently, the alcohol group is oxidized to an aldehyde and finally to a carboxylic acid, resulting in a molecule similar to a fatty acid, which can be decomposed into an acetyl-CoA by β -oxidation. This process of oxidation can occur in non-terminal carbons, giving place to two fatty acids [15]. Oxidase enzymes intervene in the metabolic routes of both alkanes such as aromatic hydrocarbons, by bacteria of the genus *Mycobacterium*, *Rhodococcus* and *Pseudomonas*, as well as by fungi of the genus *Aspergillus* and *Fusarium* [46]. In the case of polyaromatic hydrocarbons, the reaction begins with an oxidation, where two oxygenases enzyme groups participate: dioxygenases for bacteria and monooxygenases for fungi (Figure 3). In bacteria, oxygenases catalyze molecular oxygen (O_2) to the substrate, forming the intermediary cis-dihydrodiol that subsequently oxidizes, causing the rupture of the aromatic ring into muconic acids [62]. Oxidation by fungi is done by means of the P-450 Cytochrome, catalyzed by monooxygenases enzymes, to form a furan oxide (epoxide), which experiments a rearrangement to hydrolyzed phenol or a trans dihydrodiol.

Phenol is transformed into sulfates or glycosides, these compounds can originate in the detoxification of other systems that are carried out by fungi of white rot, in which the peroxidase lignin enzyme of *Phanerochaete chrysosporium*, oxidizes the dihydrodiol to form quinones, which are highly reactive compounds [46].

Anaerobic route: When the contamination is not superficial, the degradation is carried out mainly in anaerobic manner, using a variety of electron acceptors (NO_3^- , Fe^{2+} , SO_4^{2-} , CH_2) [63]. Among the microorganisms capable of degrading hydrocarbons anaerobically, the following are found: *Rhodopseudomonas palustris* CGA009 strain, *Magnetospirillum magneticum* AMB-1 strain, *Azoarcus sp.* EbN1 strain, *Geobacter metallireducens* GS-15 y *Syntrophus aciditrophicus*. Anaerobic degradation is a slower process than that of the aerobic. Additionally, it requires less energy, that is to say, the growth (and assimilation) is slower [64].

Microorganisms	HPA degraded	Enzyme	Reference
Fungi			
<i>Aspergillus ochraceae</i>	Benzo[a]pyrene	Cytochrome P450-dependent monooxygenases	[46]
<i>Penicillium spp</i>	Benzo[a]pyrene	Cytochrome P450-dependent monooxygenases	[46]
<i>Phanerochaete chrysosporium</i>	Phenanthrene, anthracene, pyrene fluoranthene	Manganese peroxidase	[47,48]
<i>Trametes versicolor</i>	Trichlorobenzene, Tetrachloroethylene	Quinone reductase, Laccase	[49]
<i>Ganoderma lucidum</i>	Benzo[a]pyrene, anthracene, Benzo[a]anthracene	Laccase	[50]
<i>Bjerkandera adusta</i>	Fluorene, anthracene	Manganese peroxidase	[51]
<i>Gleophyllum striatum</i>	Pyrene, anthracene, 9- metil anthracene, Dibenzothiophene	Lignin peroxidase	[52]
<i>Irpex lacteus</i>	Phenanthrene, anthracene, fluoranthene, pyrene	Manganese peroxidase	[53]
<i>Lentinus sp.</i>	phenanthrene, fluoranthene, pyrene	Lignin enzymes and the Cytochrome p450 monooxygenases intercellular system	[54]
Bacterium			
<i>Staphylococcus aureus</i>	Phenanthrene	Dioxygenase	[55]
<i>Mycobacterium sp. JS14</i>	Fluoranthene	Catalase and superoxide dismutase	[56]
<i>Burkholderia xenovorans</i>	Benzoate, Biphenyl	Biphenyl dioxygenase	[57]
<i>Bacillus licheniformis</i>	Naphthalene	Dioxygenases and monooxygenases.	[58]
<i>Methylococcus</i>	alkanes, alkenes and cycloalkanes (C1-C8)	Methane monooxygenases Soluble	[59]
<i>Acitobacter</i>	Alkanes and cycloalkanes (C5-C16)	Cytochrome P-450 Bacterial System	[14]
<i>Nocardia otitidiscaviarum TSH1</i>	Naphthalene	Dioxygenase	[60]

Table 1: Types of hydrocarbon decomposing microorganisms.

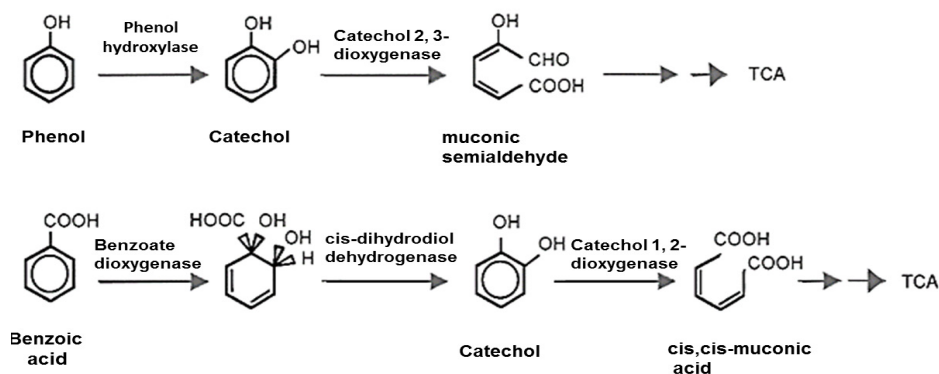


Figure 3: Phenol and benzoic acid routes in which the action of the monooxygenases (phenol hydroxylase), dioxygenases that hydroxylates rings (benzoate dioxygenase) and dioxygenase which break rings (catechol 2,3 dioxygenase). TCA, tricarboxylic acid cycle.

	Aerobic	Anaerobic
Activation of the aromatic ring	O ₂	CH ₃ , H ₂ O, COOH
Electron acceptor end	O ₂	NO ⁻³ , SO ₄ ²⁻ , Fe (III), Mn ⁺⁴ y CO ₂
Biomass production	higher production	Less production
Nutritional requirements	Less	High
Intermediate products	Catechol, protocatechuate	Benzoyl CoA, phloroglucinol, resorcinol
Enzymes	Mono, di-oxygenases	CoA ligase, Benzoyl CoA
Final products	CO ₂ , H ₂ O	CH ₄ , CO ₂ , H ₂ /N ₂

Table 2: Comparative table of the aerobic and anaerobic processes in aromatic compound degradation.

Monochromatic compound degradation: One of the best characterized routes is by toluene, where the participation of sulfate-reducing and denitrifying bacteria takes place. This also implies the addition of fumarate to toluene, followed by the activation with acetyl CoA. A benzoyl-CoA molecule is formed previously to a series of additional steps. Such molecule is susceptible to loss of aromaticity through processes of reduction, eliminating two of the three double bonds of the molecular [64].

Alkane degradation has been observed in two bacteria, a sulfate-reducing and a denitrifying (nitrate-reducing); the activation mechanism of the alkane implies the addition of a fumarate molecule, forming a diacid that is likely to be metabolized by means of oxidation of the fatty acids [17].

It has been determined that petroleum and its products degradation is quicker in the presence of oxygen than without it, this has been demonstrated in comparative studies of cultures with presence and absence of oxygen (Table 2). Although the degraded hydrocarbon spectrum in the presence of oxygen is greater, it has been observed that benzene, toluene, xylene and ethylbenzene are degraded in anaerobic conditions, and these are more soluble in water than the aromatic hydrocarbons, which make them more accessible to degradation by anaerobic microorganisms.

Conclusions

Bioremediation is considerably beneficial to the environment if it is adequately performed. To achieve this, it is crucial to develop technologies proper to the location to be decontaminated, considering previous physicochemical and microbial characterization of the site: determining with it, the concentration, the type and toxicity of the contaminant, the impacted ground area and volume, and the nutritional and environmental conditions of the site. Subsequently, it

is highly recommended to perform treatability tests in the laboratory, pilot level, to define the most appropriate treatment conditions, decide the type of process to be applied and reduce time without increasing costs for its application on a large scale.

When applying a bioremediation system, it is important to consider using the organisms (fungi, bacteria, yeast and plants) that have survived the ground contamination; because of their adaptability in contaminated ground, they are crucial candidates as bioremediators. Using their tolerance, defense and biodegradation mechanisms is a fast and efficient process that does not generate pollutants, is low in cost, and allows to restore hydrocarbon contaminated soils.

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References

- Islas-Garcia A, Vega-Loyo L, Aguilar-Lopez R (2015) Evaluation of hydrocarbons and organochlorine pesticides and their tolerant microorganisms from an agricultural soil to define its bioremediation feasibility. J Env Sci Health Part B-Pesticides; Food Contaminants and Agriculture Wastes 50: 99-108.
- Lageman R, Clarke R, Pool W (2005) Electro-reclamation, a versatile soil remediation solution. Eng Geol 77: 191-201.
- Vidalí M (2001) Bioremediation an overview. Pure Appl Chem 73: 1163-1172.
- Megharaj M, Ramakrishnan B, Venkateswarlu K, Sethunathan N, Naidu R (2011) Bioremediation approaches for organic pollutants: a critical perspective. Environ Int 37: 1362-1375.
- Roldán A, Calva G, Esparza F, Diaz M, Rodriguez R (2007) Application of solid culture amended with small amounts of raw coffee beans for the removal of petroleum hydrocarbon from a weathered contaminated soil. J Int Biodegr Biodegr 60: 35-39.
- Riojas H, Torres L, Moncada I, Balderas J, Gortares P (2010) Efectos de los surfactantes en la biorremediación de suelos contaminados con hidrocarburos. Revista Química Viva 3: 120-145.

7. Tang J, Lu X, Qing S, Wenyong Z (2012) Aging effect of petroleum hydrocarbons in soil under different attenuation conditions. *Agr Ecosyst Environ* 149: 109-117.
8. Widdel F, Rabus R (2001) Anaerobic biodegradation of saturated and aromatic hydrocarbons. *Curr Opin Biotechnol* 12: 259-276.
9. Kamada F, Abe S, Hiratsuka N, Wariishi H, Tanaka H (2002) Mineralization of aromatic compounds by brown-rot basidiomycetes - mechanisms involved in initial attack on the aromatic ring. *Microbiology* 148: 1939-1946.
10. Madigan M, Martinko J, Parker J (2003) Brock Biology of microorganisms. Pearson.
11. Xu R, Obbard JP (2004) Biodegradation of polycyclic aromatic hydrocarbons in oil-contaminated beach sediments treated with nutrient amendments. *J Environ Qual* 33: 861-867.
12. Das N, Chandran P (2011) Microbial degradation of petroleum hydrocarbon contaminants: an overview. *Biotechnol Res Int* 2011: 941810.
13. Plaza G, Lukasiak K, Wypych J, Nalecz-Jawecki G, Berry C, et al. (2008) Biodegradation of crude oil and distillation products by biosurfactant-producing bacteria. *Polish J Environ Stud* 17: 87-94.
14. Van Beilen JB, Funhoff EG (2007) Alkane hydroxylases involved in microbial alkane degradation. *Appl Microbiol Biotechnol* 74: 13-21.
15. Chávez-Gomez B, Quintero R, Esparza-García F, Mesta-Howard AM, Zavala FJ, et al. (2003) Removal of phenanthrene from soil by co-cultures of bacteria and fungi pre-grown on sugarcane bagasse pith. *Bioresour Technol* 89: 177-183.
16. Thenmozhi R, Nagasathya A, Thajuddin N (2011) Studies on biodegradation of used engine oil by consortium cultures. *Adv Environ Biol* 5: 1051-1057.
17. Carmona M, Zamarró MT, Blázquez B, Durante-Rodríguez G, Juárez JF, et al. (2009) Anaerobic catabolism of aromatic compounds: a genetic and genomic view. *Microbiol Mol Biol Rev* 73: 71-133.
18. Salinero KK, Keller K, Feil WS, Feil H, Trong S, et al. (2009) Metabolic analysis of the soil microbe *Dechloromonas aromatica* str. RCB: indications of a surprisingly complex life-style and cryptic anaerobic pathways for aromatic degradation. *BMC Genomics* 10: 351.
19. Satyanarayana T, Kulkarni S, Palande A, Deshpande M (2012) Bioremediation of petroleum hydrocarbons in soils microorganisms in environmental management. *Microbes and Environment* Ed Springer.
20. Gadd GM (2007) Geomycology: biogeochemical transformations of rocks, minerals, metals and radionuclides by fungi, bioweathering and bioremediation. *Mycol Res* 111: 3-49.
21. Harms H, Schlosser D, Wick LY (2011) Untapped potential: exploiting fungi in bioremediation of hazardous chemicals. *Nat Rev Microbiol* 9: 177-192.
22. Rahman M, Jusoh I, Husaini A, Seman I, Sing N (2014) Biodegradation and ligninolytic enzymes profiles of the newly synthesized organotin (IV)-treated non-durable tropical wood species. *J Biochem Tech* 5: 743-750.
23. Karigar CS, Rao SS (2011) Role of microbial enzymes in the bioremediation of pollutants: a review. *Enzyme Res* 2011: 805187.
24. Lei AP, Hu ZL, Wong YS, Tam NF (2007) Removal of fluoranthene and pyrene by different microalgal species. *Bioresour Technol* 98: 273-280.
25. Zheng Z, Obbard JP (2002) Polycyclic aromatic hydrocarbon removal from soil by surfactant solubilization and *Phanerochaete chrysosporium* oxidation. *J Environ Qual* 31: 1842-1847.
26. Steffen K, Hatakka A, Hofrichter M (2003) Degradation of benzo (a) pyrene by the litter-decomposing basidiomycete *Stropharia coronilla*: role of Manganese peroxidase. *Appl Environ Microbiol* 69: 3957-3964.
27. Baldrian P, Der Wiesche C, Gabriel J, Nerud F, Zadrzil F (2000) Influence of cadmium and mercury on activities of ligninolytic enzymes and degradation of polycyclic aromatic hydrocarbons by *Pleurotus ostreatus* in soil. *Appl Environ Microbiol* 66: 2471-2478.
28. Pérez J, Muñoz-Dorado J, De la Rubia T, Martínez J (2002) Biodegradation and biological treatments of cellulose, hemicellulose and lignin: an overview. *Int Microbiol* 5: 53-63.
29. Genovese M, Denaro R, Cappello S, Di Marco G, La Spada G, et al. (2008) Bioremediation of benzene, toluene, ethylbenzene, xylenes-contaminated soil: a biopile pilot experiment. *J Appl Microbiol* 105: 1694-1702.
30. Dzul-Puc JD, Esparza-García F, Barajas-Aceves M, Rodríguez-Vázquez R (2005) Benzo[a]pyrene removal from soil by *Phanerochaete chrysosporium* grown on sugarcane bagasse and pine sawdust. *Chemosphere* 58: 1-7.
31. Menezes Bento F, Oliveira Camargo FA, Okeke BC, Frankenberger WT (2005) Diversity of biosurfactant producing microorganisms isolated from soils contaminated with diesel oil. *Microbiol Res* 160: 249-255.
32. Chaillan F, Le Fleche A, Bury E, Phantavong YH, Grimont P, et al. (2004) Identification and biodegradation potential of tropical aerobic hydr.
33. Hesham A, Khan S, Tao Y, Li D, Zhang Y, et al. (2012) Biodegradation of high molecular weight PAHs using isolated yeast mixtures: application of metagenomic methods for community structure analyses. *Environ Sci Pollut Res Int* 19: 3568-3578.
34. Pan B, Xing B, Tao S, Liu W, Lin X, et al. (2007) Effect of physical forms of soil organic matter on phenanthrene sorption. *Chemosphere* 68: 1262-1269.
35. Sood N, Patle S, Lal B (2010) Bioremediation of acidic oily sludge-contaminated soil by the novel yeast strain *Candida digboiensis* TER1 ASN6. *Environ Sci Pollut Res Int* 17: 603-610.
36. Walker J, Colwell R, Vaituzis Z, Meyer S (1975) Petroleum degrading achlorophyllous alga *Prototheca zopfii*. *Nature* 254: 423-424.
37. Sivakumar G, Xu J, Thompson RW, Yang Y, Randol-Smith P, et al. (2012) Integrated green algal technology for bioremediation and biofuel. *Bioresour Technol* 107: 1-9.
38. Borde X, Guieysse B, Delgado O, Muñoz R, Hatti-Kaul R, et al. (2003) Synergistic relationships in algal-bacterial microcosms for the treatment of aromatic pollutants. *Bioresour Technol* 86: 293-300.
39. Coupe S, Smith H, Newman A, Puehmeier T (2003) Biodegradation and microbial diversity within permeable pavements. *Europ J Prot* 39: 495-498.
40. Bogus Aawska-Was E, Dabrowski W (2001) The seasonal variability of yeasts and yeast-like organisms in water and bottom sediment of the Szczecin Lagoon. *Int J Hyg Environ Health* 203: 451-458.
41. Arrieta O, Rivera A, Arias L, Rojano B, Ruiz O, et al. (2012) Biorremediación de un suelo con diésel mediante el uso de microorganismos autóctonos. *Gestión y Ambiente* 15: 27-40.
42. Taketani RG, Franco NO, Rosado AS, Van Elsas JD (2010) Microbial community response to a simulated hydrocarbon spill in mangrove sediments. *J Microbiol* 48: 7-15.
43. Mohamad F, Raja AR, Basri M (2004) Biodegradation of hydrocarbons in soil by consortium. *Int Biodeter Biodegr* 54: 61-67.
44. Simarro R, Gonzalez N, Bautista LF, Molina MC (2013) Assessment of the efficiency of in situ bioremediation techniques in a creosote polluted soil: change in bacterial community. *J Hazard Mater* 262: 158-167.
45. Chen M, Xua P, Zenga G, Yang C, Huang D, et al. (2015) Bioremediation of soils contaminated with polycyclic aromatic hydrocarbons, petroleum, pesticides, chlorophenols and heavy metals by composting: Applications, microbes and future research needs. *Biotech Adv* 33: 745-755.
46. Romero MC, Salvioli ML, Cazau MC, Arambarri AM (2002) Pyrene degradation by yeasts and filamentous fungi. *Environ Pollut* 117: 159-163.
47. Cortes D, Absalon A (2013) Phenanthrene removal from soil by a strain of *Aspergillus niger* producing Manganese peroxidase of *Phanerochaete chrysosporium* Chemistry "Hydrocarbon". Kutchurov V, Kolesnikov A.
48. Bishnoi K, Kumar R, Bishnoi N (2008) Biodegradation of polycyclic aromatic hydrocarbons by white rot fungi *Phanerochaete chrysosporium* in sterile and unsterile soil. *Int J Recent Sci Res* 67: 538-542.
49. Marco-Urrea E, Perez-Trujillo M, Caminal G, Vicent T (2009) Dechlorination of 1,2,3- and 1,2,4-trichlorobenzene by the white-rot fungus *Trametes versicolor*. *J Hazard Mater* 166: 1141-1147.
50. Punnapayak H, Prasongsuk S, Messner K, Danmek K, Lotrakul P (2009) Polycyclic aromatic hydrocarbons (PAHs) degradation by laccase from a tropical white rot fungus *Ganoderma lucidum*. *Afr J Biotechnol* 8: 5897-5900.
51. Haritash AK, Kaushik CP (2009) Biodegradation aspects of polycyclic aromatic hydrocarbons (PAHs): a review. *J Hazard Mater* 169: 1-15.
52. Yadav M, Singh S, Sharma J, Deo Singh K (2011) Oxidation of polyaromatic hydrocarbons in systems containing water miscible organic solvents by the lignin peroxidase of *Gleophyllum striatum* MTCC-1117. *Environ Technol* 32: 1287-1294.

53. Baborova P, Moder M, Baldrian P, Cajthamlová K, Cajthamí T (2006) Purification of a new manganese peroxidase of the white-rot fungus *Irpex lacteus*, and degradation of polycyclic aromatic hydrocarbons by the enzyme. *Res Microbiol* 157: 248-253.
54. Juckpech K, Onruthai P, Panan R (2012) Degradation of polycyclic aromatic hydrocarbons by newly isolated *Curvularia* sp. F18, *Lentinus* sp. S5, and *Phanerochaete* sp. T20. *Science Asia* 38: 147-156.
55. Mallick S, Chatterjee S, Dutta TK (2007) A novel degradation pathway in the assimilation of phenanthrene by *Staphylococcus* sp. strain PN/Y via meta-cleavage of 2-hydroxy-1-naphthoic acid: formation of trans-2,3-dioxo-5-(2'-hydroxyphenyl)-pent-4-enoic acid. *Microbiology* 153: 2104-2115.
56. Lee SE, Seo JS, Keum YS, Lee KJ, Li QX (2007) Fluoranthene metabolism and associated proteins in *Mycobacterium* sp. JS14. *Proteomics* 7: 2059-2069.
57. Parnell JJ, Park J, Denev V, Tsoi T, Hashsham S, et al. (2006) Coping with polychlorinated biphenyl (PCB) toxicity: Physiological and genome-wide responses of *Burkholderia xenovorans* LB400 to PCB-mediated stress. *Appl Environ Microbiol* 72: 6607-6614.
58. Lin C, Gan L, Chen ZL (2010) Biodegradation of naphthalene by strain *Bacillus fusiformis* (BFN). *J Hazard Mater* 182: 771-777.
59. McDonald I, Miguez C, Rogge G, Bourque D, Wendlandt K, et al. (2006) Diversity of soluble methane monooxygenase-containing methanotrophs isolated from polluted environments. *FEMS Microbiol Lett* 255: 225-232.
60. Zeinali M, Vossoughi M, Ardestani SK (2008) Naphthalene metabolism in *Nocardia otitidiscaviarum* strain TSH1, a moderately thermophilic microorganism. *Chemosphere* 72: 905-909.
61. Supaphol S, Panichsakpatana S, Trakulnaleamsai S, Tungkananuruk N, Roughjanajirapa P, et al. (2006) The selection of mixed microbial inocula in environmental biotechnology: example using petroleum contaminated tropical soils. *J Microbiol Methods* 65: 432-441.
62. Cerniglia CE (1997) Fungal metabolism of polycyclic aromatic hydrocarbons: past, present and future applications in bioremediation. *J Ind Microbiol Biotechnol* 19: 324-333.
63. Akpor O, Igbinosa OE, Igbinosa O (2007) Studies on the effect of petroleum hydrocarbon on the microbial and physico-chemicals characteristics of soil. *Afr J Biotechnol* 6: 1939-1943.
64. Peters F, Shinoda Y, McInerney J, Boll M (2007) Cyclohexa-1,5-diene-1-carbonyl-coenzyme A (CoA) hydratases of *Geobacter metallireducens* and *Syntrophus aciditrophicus*: evidence for a common benzoyl-CoA degradation pathway in facultative and strict anaerobes. *J Bacteriol* 189: 1055-1060.