

# Arsenic Occurrence and Fate in the Environment; A Geochemical Perspective

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# Abstract

The arsenic minerals in the environment constitute the primary source of the environmentally occurring arsenic. The As minerals interact with the environment and this renders either their dissolution or the formation of secondary minerals, or both. The distribution of the environmental arsenic is determined by the biogeochemical transformations with respect to the redox conditions, the pH, the availability of ions, the adsorption-desorption, dissolution and the biological activity. The arsenic in the environment is sorbed primarily by metal oxides, especially the ones of iron (Fe), aluminum (AI) and manganese (Mn). These are thought to bind As(+5) readily than As(+3). The overall adsorption depends greatly on pH. Metal oxides such as the ones of hydrous ferric, manganese and aluminum are additional important sinks of arsenic that is adsorbed or co-precipitated. Their dissolution depends also on pH. The redox potential and the microbial activity. Final result is the release of arsenic chemical species into the environment.

This review presents a systematic compilation of the major geochemical processes that govern arsenic fate in the environment. The paper attempts to compile the removal capacity of constituents which could be useful for the purpose of As remediation.

**Keywords:** Arsenic environmental fate; Arsenic speciation; Arsenic geochemistry

## Introduction

Arsenic (As) is a trace inorganic element. It is found in the crystal lattice of arsenic minerals inspected in the environment. It may also arise from arsenic traces adsorbed onto the surfaces of -other minerals as inner or outer sphere complexes. Arsenic minerals–water interaction govern the arsenic occurrence in water systems. Due to this, arsenic is traced in surface and ground waters, -for example in rivers, stream, sea waters and lakes. The rate of interaction between water and arsenic minerals depends on several ad-hoc biogeochemical conditions such as the redox potential (Eh), the pH, the temperature, the microbial activity, the speciation and the concentration of metals in the fluid and the ionic strength of the solution. Once arsenic is released it can be sorbed onto the minerals phase, precipitate, redissolved and bio-integrate according to the surrounding environmental conditions.

Arsenic pollution has become a significant environmental problem nowadays. Arsenic is a contaminant of concern regarding many industrial products, wastes and wastewaters and is widely known for its adverse effects on human health. Great human populations have been affected by the adverse health effects of arsenic [1,2]. Recent epidemiological studies have reported noteworthy detrimental effects of As on humans due to its high toxicity. The toxicity occurs even at the ppb level [1,2]. Due to the above reasons, the World Health Organization (WHO) suggested that the arsenic concentration in drinking water should not exceed 10 µg/L [1]. Note, that the upper concentration limit of As in drinking water was 50 µg/L [1]. Arsenic is a known neurotoxin [2]. There are several studies which link arsenic exposure to neuropathological disorders such as production of ß amyloid [3], hyperphosphorylation of tau protein [4], oxidative stress [5], inflammation [6], endothelial cell dysfunction [7], and angiogenesis [8]. Note that some of these disorders are related to cognitive dysfunction and Alzheimer's disease [9]. Morphologic and neurochemical alterations have also been observed in animals during arsenic exposure. These have been associated with the hippocampus and other memory-related neuronal structures related learning and memory [2,7]. Skin diseases and carcinogenesis are other effects observed due to the arsenic exposure according to human epidemiological studies [10]. Most importantly, considerably high were the estimated cancer risks for wide population groups which were exposed to arsenic-contaminated water in Taiwan, Japan, Bangladesh, West Bengal-India, Chile and Argentina [11]. The higher risks were identified for cancer of the skin and then for cancers of lung, bladder, kidney and liver [11].

Nowadays several soil and groundwater reservoirs have been contaminated with arsenic. This has created the need for remedy against arsenic contamination. However, the treatment and remediation of As is not an easy task to accomplish since arsenic changes several valence states and reacts towards the formation of species with varying toxicity and mobility. It is this reason that makes the removal of arsenic from drinking water a really recurring challenge, especially in the developing countries. It should be noted that there are several reasons apart from cost that can make it advantageous to use local materials as adsorbents for arsenic. Such materials can be retrieved from the nearby soils as well as the topical agriculture and industry.

Of course these materials are not always optimal, but their availability at various locations all over the world often makes them attractive choices.

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#### Arsenic Origin in the Environment

Traces of arsenic can be found in several parts of the natural environment. Arsenic is traced in air, soil, water, rocks, plants and animals. There are various natural activities that release arsenic to the environment. Most significant are the volcanic eruption, the erosion of the rocks and the forest fires. Arsenic is widely distributed in more than 320 minerals [12]. The most common arsenic minerals are:

- (1) arsenopyrite (FeAsS) [13-15];
- (2) orpiment  $(As_2S_3)$ ;
- (3) realgar ( $As_2S_2$ );
- (4) pyrite (FeS<sub>2</sub>) where arsenic is found as a solid solution [16,17].

After weathering of arsenic minerals the arsenic ions can be adsorbed onto the Fe (III) and Mn (IV) oxide-hydroxide phases [15,18-20]. The arsenic bearing minerals are also significant sources of arsenic present in natural water. Apart from that, traces of As in natural water are observed due to the widespread use of [arsenic in pigments, insecticides and herbicides. Note that approximately 70% of the arsenic used, is in pesticides. In the form of pesticides it is principally traced in [21]:

- (1) Monosodium methane arsenate (MSMA) HAsO<sub>3</sub>CH<sub>3</sub>Na;
- (2) Disodium methane arsenate (DSMA) Na2AsO3CH3 and
- (3) Dimethylarsinic acid (cacodylic acid) (CH<sub>3</sub>)<sub>2</sub>AsO<sub>2</sub>H;
- (4) Arsenic acid  $H_3AsO_4$

In general, variable and low is the concentration of the arsenic in the environment [22]. For example, the average concentration in air in remote and rural areas ranges between 0.02 ngm<sup>-3</sup> and 4 ngm<sup>-3</sup> [22]. In urban areas the ambient concentration of arsenic is from 3 ngm<sup>-3</sup> to about 200 ngm<sup>-3</sup>. High values, above 1000 ngm<sup>-3</sup>, can be measured near industries. In ocean waters arsenic concentrates typically between 1  $\mu$ gl<sup>-1</sup> and 2  $\mu$ gl [22]. Arsenic is also widely distributed in surface fresh waters, rivers and lakes. Note that concentrations below 10  $\mu$ gl<sup>-1</sup> are usually addressed in rivers and lakes, however certain water samples may present arsenic concentration up to 5 mgl<sup>-1</sup>, especially near anthropogenic sources [22]. The levels of arsenic in groundwater average to about 1-2  $\mu$ gl<sup>-1</sup>. Worth to mention is that in areas with volcanic rock and sulfide mineral deposits, the arsenic levels may rise up to 3 mg l<sup>-1</sup> [22].

There are numerous regions in the planet where the arsenic concentrations exceed the newly suggested Maximum Contaminant Level (MCL=10  $\mu$ gl<sup>-1</sup>) [1]. For example, waters in U.S., India and China, exceeds MCL value. It is notable that in Bangladesh about 30% of the ground water sources, have arsenic concentrations above the MCL concentration limit, exceeds 50  $\mu$ gl<sup>-1</sup> [1]. In U.S., approximately 10% of the measured borehole water samples exceed 10  $\mu$ gl<sup>-1</sup> [23]. Extremely high arsenic concentrations have been reported in water samples of Xinjiang (China), where the corresponding arsenic concentrations were all well above 50  $\mu$ gl<sup>-1</sup> and more specifically between 50  $\mu$ gl<sup>-1</sup> and 1860  $\mu$ gl<sup>-1</sup> [24]. Another disconcerting issue is that high arsenic bioaccumulation rates in the dry weight range of 0.007  $\mu$ gg<sup>-1</sup> and 125.9  $\mu$ gg<sup>-1</sup> have been reported, as a result of its gathering into the tissues of aquatic biota [25]. Table 1 presents the major arsenic sources in earth materials.

The concentration of arsenic in sedimentary materials range from 1.7 to 400 mgkg<sup>-1</sup> [26]. Higher levels occur in contaminated areas. The background concentration of arsenic in soil is between 1 mgkg<sup>-1</sup> and

Arsenic Minerals	Chemical formula	Source	Reference
Native arsenic	As	Hydrothermal veins.	[1]
Niccolite	NiAs	Vein deposits and norites.	[1]
Realgar	AsS	Vein deposits, often associated with orpiment, clays andlimestones, also deposits from hot springs.	[1]
Orpiment	As <sub>2</sub> S <sub>3</sub>	Hydrothermal veins, hot springs, volcanic sublimationproducts.	[1]
Cobaltite	CoAsS	High temperature deposits, metamorphic rocks.	[1]
Arsenopyrite	FeAsS	The most abundant As mineral, dominantly in mineral veins.	[1]
Tennantite	(Cu,Fe) <sub>12</sub> As <sub>4</sub> S <sub>13</sub>	Hydrothermal veins.	[1]
Enargite	Cu <sub>3</sub> AsS <sub>4</sub>	Hydrothermal veins.	[1]
Arsenolite	As <sub>2</sub> O <sub>3</sub>	Secondary mineral formed by oxidation of arsenopyritenative arsenic and other As minerals.	[1]
Clauderite	As <sub>2</sub> O <sub>3</sub>	Secondary mineral formed by oxidation of realgararsenopyrite and other As minerals.	[1]
Scorodite	FeAsO <sub>4</sub> •2H <sub>2</sub> O	Secondary mineral.	[1]
Anabergite	(Ni,Co) <sub>3</sub> (AsO <sub>4</sub> ) <sub>2</sub> •8H <sub>2</sub> O	Secondary mineral.	[1]
Hoernesite	Mg <sub>3</sub> (AsO <sub>4</sub> ) <sub>2</sub> •8H <sub>2</sub> O	Secondary mineral, smelter wastes.	[1]
Haematolite	(Mn,Mg) <sub>4</sub> Al(AsO <sub>4</sub> )(OH) <sub>8</sub>	Secondary mineral.	[1]
Conichalcite	CaCu(AsO <sub>4</sub> )(OH)	Secondary mineral.	[1]
Pharmacosiderite	Fe <sub>3</sub> (AsO <sub>4</sub> ) <sub>2</sub> (OH) <sub>3</sub> •5H <sub>2</sub> O	Oxidation product of arsenopyrite and other As minerals.	[1]
Materials	Concentration As (mg/kg)	Process	
Igneous material		Cooling and solidification of magma or lava.	[26]
Basalt	<1-113		[26]
Ultrabasics	<1-16		[26]
Granites	<1-15		[26]
Sedimentary material		Formed by the deposition of material (organic and/orminerals) at the Earth's surface and within bodies of water.	[26]
Shales and clays	<1-500		[26]
Sandstones	<1-120		[26]
Limestones	<1-20		[26]
Phosphorites	3-100		[26]

Table 1: Arsenic sources and concentrations in earth materials; adopted and modified from [1,26,27].

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40 mg kg<sup>-1</sup>, with a mean value around 5 mg kg<sup>-1</sup> [27]. Significant is that near industries, mines and mine tailings, the arsenic concentration in the soil and soil sediments is much higher [1]. For example, an average arsenic concentration of 903 mg/kg-1 was detected in mine tailings in British Columbia [28]. On the other hand, the marine organisms contain arsenic residues typically from 1 mg·kg<sup>-1</sup> to about 100 mg·kg<sup>-1</sup>. Predominantly higher arsenic concentrations are addressed in organic arsenic species such as the arsenosugars (macroalgae) and the arsenobetaine (invertebrates and fish) [22]. Various anthropogenic activities act as additional sources of arsenic in the environment as for example farming, mining, uses of fossil fuels, pulp and paper production and cement manufacturing [29]. Other significant anthropogenic activities contributes to an input of Table 1-arsenic and its compounds in the environment are wood preservatives, glass manufacture, electronics, catalysts, alloys, feed additives and veterinary chemicals.

## Fate of Arsenic in Nature

Since the primary source of arsenic in the natural environment is the arsenic minerals, the stability of these is a significant factor of controlling arsenic occurrence in nature. The interaction of the arsenic minerals with the environment results either to their dissolution, or to the formation of secondary arsenic minerals, or even both. Table 2, presents characteristic cases of solubility data of representative arsenic minerals and the Gibbs free energy of formation in their standard reference state ( $_{f}G^{0}$ ) [reference 30 and references therein]. Note that the values of pH and the corresponding total arsenic solubility of Table 2, are the highest and lowest levels presented in the literature (Table 2).

The most common valence states of arsenic in natural systems are the +3 and +5 states. However, arsenic can be found in the (-3) oxidation state (arsine), and an arsenic compound in this state is extremely toxic. This toxic oxidation state can be formed under very reducing conditions, and for this reason its occurrence in nature is relatively rare. On the other hand, inorganic and organic species of As are present in the natural environment, with inorganic forms been typically more abundant in freshwater systems.

In aqueous systems, arsenic exhibits anionic behaviour. In case of oxygenated waters, arsenic acid predominates only at extremely low pH values, namely for pH below 2. In the pH range of 2 to 11, it is in the form of  $H_2AsO_4^-$  and  $HAsO_4^{2-}$ . In mildly reduced conditions and low pH values, arsenious acid is converted to  $H_2AsO_3^-$  and this conversion is more frequent as pH increases. When the pH exceeds 12  $HAsO_3^{2-}$  does appear (Figure 1).

Arsenate  $[As(+5); HxAsO_4^{x\cdot3}, x=0-3]$  and arsenite  $[As(+3); HxAsO_3^{x\cdot3}, x=0-3]$  are the two most common inorganic forms of arsenic in freshwaters. As(+5) is thermodynamically stable under oxic conditions, while As(+3) is stable under more reducing conditions. However, As(+5) and As(+3) are often found in both oxic and anoxic waters and sediments. The oxidation of As(+3) by O<sub>2</sub> is slow (on the order of several weeks), while bacterially-mediated redox reactions can be much faster [30-33].

Arsenate is an anion at the pH of most natural waters ( $H_2AsO_4^-$  and  $HAsO_4^{-2}$ ), while arsenite is a neutral specie. The pKa values for arsenate ( $H_3AsO_4$ ) are pKa1=2.19, pKa2=6.94 and pKa3=11.5 according to the following equations (1-4):

$$H_{2}AsO_{4} = H_{2}AsO_{4}^{-} + H^{+}$$

$$\tag{1}$$

$$H_2AsO_4^- = HAsO_4^{-2} + H^+$$
(2)

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$$HAsO_4^{-2} = AsO_4^{-3} + H^+$$
(3)

$$H_{3}AsO_{3} = H_{2}AsO_{3}^{-} + H^{+}$$
(4)

In oxidative environments the form  $H_2AsO_4^-$  predominates with pH values below 6.9, whereas the HAsO<sub>4</sub><sup>-2</sup> ions predominate at higher pH levels. Arsenite (H<sub>3</sub>AsO<sub>3</sub>) pKa value according to equation (4) is equal to 9.22, while it is the main arsenic chemical specie in natural waters with pH<9 and in slightly reducing conditions [34].

In aerobic waters, arsenic acid predominates only at extremely low pH (<2). At the pH range of 2 to 11, it is replaced by  $H_2AsO_4^-$  and  $HAsO_4^{2-}$  ions. Arsenious acid appears at low pH and under mildly reduced conditions, but it is replaced by  $H_2AsO_3^-$  as the pH increases. Only when the pH exceeds 12 the  $HAsO_3^{2-}$  ion appears (Figure 1). The  $HAsS_2$  arsenic chemical specie can form at low pH in the presence of sulphide ions. Arsine derivatives and arsenic metal can occur under extreme reducing conditions [35].

Arsenic changes its valence state and chemical form in the environment. In the pH range of 4 to 10, As(+5) species are negatively charged in water, and the predominant As(+3) species is neutral in charge (Figure 1).

The pH, the redox potential and the arsenic speciation are the critical parameters for the Eh-pH diagrams as those of Figure 1. If the half-reaction for the reduction of As(+5) to As(+3) is accounted in equation (5), the Nernst equation (6) relates the arsenic species concentrations at equilibrium [36], namely:

$$H_3AsO_4 + 2H^+ + 2e^- = H_3AsO_3 + H_2O$$
(5)

$$E = E^{0} + k \left( Log_{10} \left( \frac{H_{3}AsO_{4}}{H_{3}AsO_{3}} \right) - 2pH \right)$$
(6)

In equation 6,  $E^0$  and k are a constant and a collection of constants, respectively. The dependence of the As(+5) fraction with the pH of the studied aquatic system is given by equation (7) as proposed by Stumm and Morgan 1996 [37]:

$$a^{+5} = \frac{\left[H_3 A s O_4\right]}{A s (+5)} = \frac{1}{1 + K_{a1} 10^{pH} + K_{a1} K_{a2} 10^{2pH} + K_{a1} K_{a2} K_{a3} 10^{3pH}}$$
(7)



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Mineral	Formula	Crystal system	Arsenic solubility conditionsa	$\Delta fG^0$
			-	(kJ·mol⁻¹)
Arsenolite	As <sub>2</sub> O <sub>3</sub>	Cubic	11.1 g/L; 22°C	-576.34
Claudetite	As <sub>2</sub> O <sub>3</sub>	Monoclinic	10.1 g/L; 22°C	-576.53
AFA/pitticite	$Fex(AsO_4)y(SO_4)z \cdot nH_2O$	Amorphous	75-15370 mg/L at pH 0.5-2.4; 25-130 mg/L at pH 1.82-3.10; 25°C	-1268.72; -1267.1
Arseniosiderite	Ca <sub>2</sub> Fe <sub>3</sub> O <sub>2</sub> (AsO <sub>4</sub> ) <sub>3</sub> •3H <sub>2</sub> O	Monoclinic	3.1-27 mg/L at pH 6.85-8.15; 25°C	-
Kaatialaite	$Fe(H_2AsO_4)_3 \bullet 5(H_2O)$	Monoclinic	5.4 g/L; TCLPb test	-
Kaňkite	Fe <sub>3</sub> (AsO <sub>4</sub> )•3.5H <sub>2</sub> O	Monoclinic	-	-
Kolfanite	Ca <sub>2</sub> Fe <sub>3</sub> O <sub>2</sub> (AsO <sub>4</sub> ) <sub>3</sub> •2H <sub>2</sub> O	Monoclinic	1.36 mg/L at pH 8; 20°C	-
Parasymplesite	Fe <sub>3</sub> (AsO <sub>4</sub> )•8H <sub>2</sub> O	Monoclinic	-	-
Pharmacosiderite	K[Fe <sub>4</sub> (OH) <sub>4</sub> (AsO <sub>4</sub> ) <sub>3</sub> ]•6.5H <sub>2</sub> O	Cubic	-	-
Scorodite	FeAsO <sub>4</sub> •2H <sub>2</sub> O	Orthorombic	0.33-5.89 mg/L at pH 5.01-6.99; 22°C; 0.11-463 mg/L at pH 0.97-7.92; 23°C; 1.8-10.3 mg/L at pH 5.53-6.36; 25°C	-1282.42; -1285.05; -1279.2; -1263.52
Symplesite	$Fe_3(AsO_4)_2 \cdot 8H_2O$	Triclinic	0.024-7 mg/L at pH 6.0-9.1; 27°C	-3751.02; -3792.01
Yukonite	Ca <sub>7</sub> Fe <sub>12</sub> (AsO <sub>4</sub> ) <sub>10</sub> (OH) <sub>20</sub> •15H <sub>2</sub> O	Amorphous	1.16-5.11 at pH 7.56-8.82; 20°C; 6.3-51 mg/L at pH 5.5-6.15; 25°C	-
Beudantite	PbFe <sub>3</sub> (AsO <sub>4</sub> )(SO <sub>4</sub> )(OH) <sub>6</sub>	Hexagonal	<0.02 mg/L at pH 4.3-4.65; 25°C	-3055.6; -3081.12
Bukovskýite	Fe <sub>2</sub> (AsO <sub>4</sub> )(SO <sub>4</sub> )(OH)•7H <sub>2</sub> O	Triclinic	-	-3480
Sarmientite	Fe <sub>2</sub> (AsO <sub>4</sub> )(SO <sub>4</sub> )(OH)•5H <sub>2</sub> O	Monoclinic	-	-
Tooeleite	Fe <sub>6</sub> (AsO <sub>3</sub> ) <sub>4</sub> (SO <sub>4</sub> )(OH) <sub>4</sub> •4H <sub>2</sub> O	Monoclinic	-	-
Zýkaite	Fe <sub>4</sub> (AsO <sub>4</sub> ) <sub>3</sub> (SO <sub>4</sub> )(OH)•15H <sub>2</sub> O	Orthorombic	-	-
Haidingerite	Ca(AsO <sub>3</sub> OH)•H <sub>2</sub> O	Orthorombic	2050 mg/L at pH 6.22; 23°C 3120–4360 mg/L at pH 4.93; TCLPb test	-1533
Hörnesite	$Mg_3(AsO_4)_2 \cdot 8H_2O$	Monoclinic	300-1100 mg/L at pH 6.5-7.4	-
Pharmacolite	Ca(HAsO <sub>4</sub> )•2H <sub>2</sub> O	Monoclinic	5919 mg/L at pH 6.7; 25°C 3120-4360 mg/L at pH 4.93; TCLPb test	-1808.21
Picropharmacolite	Ca <sub>4</sub> Mg(AsO <sub>4</sub> ) <sub>2</sub> (HAsO <sub>3</sub> OH) <sub>2</sub> •11H <sub>2</sub> O	Triclinic	-	-
Weilite	CaHAsO₄	Triclinic	2170-3610 mg/L at pH 4.93; TCLPb test 540-764 mg/L at Ph 3-8; 35°C	-1292.48
Adamite	Zn <sub>2</sub> (AsO <sub>4</sub> )(OH)		-	-1252.29
Annabergite	Ni <sub>3</sub> (AsO <sub>4</sub> ) <sub>2</sub> •8H <sub>2</sub> O	Orthorombic	47.8-1449 mg/L at pH 3-9; 22°C	-3488.57; -3482.34
Austinite	CaZn(AsO <sub>4</sub> )(OH)	Orthorombic	-	-1651.13
Bayldonite	PbCu <sub>3</sub> (AsO <sub>4</sub> ) <sub>2</sub> (OH) <sub>2</sub>	Triclinic	-	-1810.6
Clinoclase	Cu <sub>3</sub> (AsO <sub>4</sub> )(OH) <sub>3</sub>	Monoclinic	-	-1209.48
Conichalcite	CaCu(AsO <sub>4</sub> )(OH)	Orthorombic	-	-1470.17
Cornubite	Cu <sub>5</sub> (AsO <sub>4</sub> ) <sub>2</sub> (OH) <sub>4</sub>	Triclinic	-	-2057.9
Duftite	PbCu(AsO₄)(OH)	Orthorombic	-	-959.92
Erythrite	Co <sub>3</sub> (AsO <sub>4</sub> ) <sub>2</sub> •8H <sub>2</sub> O	Monoclinic	-	-
Euchroite	Cu <sub>2</sub> (AsO <sub>4</sub> )(OH)•3(H <sub>2</sub> O)	Orthorombic	-	-1552.7
Fornacite	Pb <sub>2</sub> Cu(AsO <sub>4</sub> )(CrO <sub>4</sub> )(OH)	Monoclinic	0.03 mg/L at pH 6.96	-1956.86l
Köttigite	Zn <sub>3</sub> (AsO <sub>4</sub> ) <sub>2</sub> •8H <sub>2</sub> O	Monoclinic	16 mg/L at pH 4.87	-4030.48
Legrandite	Zn <sub>2</sub> (AsO <sub>4</sub> )(OH)•H <sub>2</sub> O	Monoclinic	-	-1488.6
Mansfieldite	AIAsO <sub>4</sub> •2H <sub>2</sub> O	Orthorombic	-	-1730.78; -1720.8
Mimetite	Pb <sub>5</sub> (AsO <sub>4</sub> ) <sub>3</sub> Cl	Hexagonal	-	–2675.5; –2616.8
Olivenite	Cu <sub>2</sub> (AsO <sub>4</sub> )(OH)	Monoclinic	-	-845.52
Schultenite	Pb(AsO <sub>3</sub> OH)	Monoclinic	8.8 mg/L at pH 4.68; 25°C	-805.66; -809.62
Sterlinghillite	Mn <sub>3</sub> (AsO <sub>4</sub> ) <sub>2</sub> •4H <sub>2</sub> O	Monoclinic	-	-4045.17
Arsenopyrite	FeAsS	Monoclinic	-	-141.6
α,Realgar	AsS	Monoclinic	-	-31.3
β,Realgar	AsS	Monoclinic	-	-30.9
α,Orpiment	As <sub>2</sub> S <sub>3</sub>		-	-84.9
am, Orpiment	As <sub>2</sub> S <sub>3</sub>	Monoclinic	-	-76.8

a.: pH and total arsenic solubility values are the highest and lowest levels presented and there is no relation between solubility and pH values.

b.: TCLP means Toxicity Characteristic Leaching Procedure.

Table 2: Arsenic minerals formula, crystal system, ΔfG<sup>0</sup> and solubility for standard state conditions, 298.15 K and 1 atm; adopted and modified from [27,30,31].

A similar equation can be derived for the fraction of As(+3) in the  $H_3AsO_3$  form ( $\alpha^{+3}$ ) due to equation (8):

Substitution of equations (7) and (8) into equation (6) give equation (9), which relates the equilibrium redox potential to the pH and concentrations of As(+5) and As(+3) measured quantities:

$$a^{+3} = \frac{\left[H_3 A s O_3\right]}{A s (+3)} = \frac{1}{1 + K_{a1} 10^{pH} + K_{a1} K_{a2} 10^{2pH} + K_{a1} K_{a2} K_{a3} 10^{3pH}}$$
(8)

$$E = E^{0} + k \left( Log_{10} \left( \frac{As(+5) a^{+5}}{As(+3) a^{+3}} \right) - 2pH \right)$$
(9)

The major conditions responsible for different arsenic valance and chemical species are redox potential, the presence of complexing ions, such as ions of sulfur, iron, calcium and microbial activity. Since arsenic forms anions in solution, it does not form complexes with simple anions like Cl<sup>-</sup> and SO<sub>4</sub><sup>2-</sup> as do cationic metals, whereas anionic arsenic complexes behave like ligands in water.

## Arsenic Geochemistry

The geochemical processes which are involved in the oxidation, reduction and bioavailability of chemical species of arsenic play a significant role in nature [34]. Arsenic has been associated to microorganisms as well and especially their metabolism [34]. Arsenic is partitioned between solid and dissolved phases and this is associated with many biogeochemical processes. There are also several reactions that control the mobilization of arsenic in the natural environment. Most significant are the processes of dissolution-precipitation, adsorption-co precipitation and reduction-oxidation.

Oxidizing arsenic bacteria, oxidize As(+3) enzymatically and produce arsenite oxidases. Oxygen and nitrate reducing microorganisms, utilize arsenite as an electron donor. Characteristic examples are the photoautotrophic, heterotrophic and chemoautotrophic microorganisms. In these micro-organisms, the adduced energy is used for the production of CO<sub>2</sub> which is required for the generation of carbon and the growth of bacteria community [34,38]. The oxidation of the As(+3) species to the less bioavailable As(+5) compounds is crucial for the detoxification processes [34,38]. Such processes can be observed in extreme natural environments [34,38]. They are considered as primary energy resources for the chemolithotrophic metabolism of organisms in the era of the formation of the very first forms of life [34,38]. The microorganisms which can act as oxidizers of arsenite, can also facilitate As(+3) oxidation in aerobic environments [37], viz. in the presence of O<sub>2</sub> They can also act as electron acceptors in anoxic conditions by using other ions in the order of NO,  $\rightarrow$  Mn oxides,  $\rightarrow$  Fe(III) oxides,  $\rightarrow$  sulfate [37]. On the contrary, in the case of reducing bacteria, the process is the reduction of As(+5) to As(+3). Note that this process is related to the detoxification of the cells. The arsenate ions enter the cells via the phosphate transporters (for example Pst - high affinity phosphate specific transport or Pit - low affinity phosphate inorganic transport), due to structural homologies with phosphate ions. After reaching the cytoplasm, arsenate is reduced into arsenite. This is accomplished by the arsenate-reductase enzyme ArsC or the ArsAB complex. Note that the reduction of arsenate to arsenite is implemented before it is excreted from the cell by the transmembrane protein ArsB. It is mentioned here that ArsB is also known as Acr3 in the context of some eukaryotic micro-organisms). The transformation process followed by the excretion of arsenic is a common occurrence in the living world and is widespread in bacteria [34,39]. Nevertheless, the mobility of arsenic in the natural environment is determined generally by the extent to which it is adsorbed onto the mineral surfaces. Metal oxides tend to be the primary sorbents of As in the environment, especially the oxides of iron (Fe) and aluminum (Al). In addition Mn-oxides can also sorb arsenic to some extent. Among the most important oxides-sinks in nature for the adsorbed, co-precipitated arsenic or even both, are the hydrous ferric, manganese and aluminum oxides. These oxides are composed of octahedrally coordinated metal atoms that share edges to form chains in two and three dimensions structures. However, according to different pH values the OH groups can bind or release H<sup>+</sup> ions and this results in the development of a surface charge. In this case arsenic adsorbs this charge by the process of ligand exchange with OH and  $OH_2^+$  surface functional groups. This yield to the formation of an inner-sphere complex. This type of adsorption requires an incompletely dissociated acid, e.g.,  $H_2AsO_4^-$ , to provide a proton for the complexation process with the surface OH group so as to form  $H_2O$  and to providing a space for the anion. The reactions which delineate these geochemical processes are given below [40]:

$$\equiv SurfOH + H_3AsO_4^0 \rightarrow \equiv SurfH_2AsO_4^0 + H_2O$$
(10)  
$$\equiv SurfOH + H_2AsO_4^- \rightarrow \equiv SurfHAsO_4^- + H_2O$$
(11)

$$\equiv SurfOH + HAsO_{*}^{-2} \rightarrow \equiv SurfAsO_{*}^{-2} + H_{2}O$$
(12)

$$\equiv SurfOH + H_3AsO_4^0 \rightarrow \equiv SurfH_2AsO_3^0 + H_2O$$
(13)

$$\equiv SurfOH + H_2AsO_3^- \rightarrow \equiv SurfHAsO_3^- + H_2O \tag{14}$$

$$\equiv SurfOH + HAsO_3^{-2} \rightarrow \equiv SurfAsO_3^{-2} + H_2O$$
(15)

where,  $\equiv$ *SurfOH* represents the structural metal atom and associated OH surface functional group. For example  $\equiv$ *SurfHAsO*<sub>4</sub><sup>-</sup> is the surface arsenic complex. The energy required to dissociate the weak acid at the oxide surface and the amount of arsenic adsorbed varies with pH.

In order to describe adsorption-desorption reactions at the mineral surfaces, two types of models have been proposed. The empirical models are based on the partitioning relationships of a solute between the aqueous and the solid phase, and the conceptual models. The latter models account for the surface complexation and treat adsorption reactions similar to ion association reactions in solution. All models assume that the adsorption reactions are at equilibrium. In the case of the empirical adsorption models the distribution coefficient as in equation (16) is the most known [41]:

$$\overline{C} = K_d C \tag{16}$$

In addition the Langmuir Isotherm and the Freundlich Isotherm are given in equations (17) and (18) respectively [41].

$$\overline{C} = \frac{K_L SC}{1 + K_L C} \tag{17}$$

$$\overline{C} = K_F C^a \tag{18}$$

The  $\overline{C}$  is the adsorbed concentration, *C* is the aqueous concentration at equilibrium with the solid phase, *S* is the adsorption site concentration,  $K_d$  is the distribution coefficient,  $K_L$  is the Langmuir equilibrium constant,  $K_F$  is the Freundlich equilibrium constant, and *a* is the Freundlich exponent.

Arsenic is absorbed by several clay minerals. The maximum adsorption of As(+5) is kaolinite, montmorillonite, illite, halloysite, and chlorite for pH values near 7. The adsorption is decreased with further increase of pH above 7. Adsorption of As(+3) by the same clay minerals is low at low pH values and is increased when pH increases. Arsenate is adsorbed to a greater extent than As(+3) on all clay minerals at pH<7. At higher pH values, adsorption of As(+5) and As(+3) are more comparable, and in some cases As(+3) adsorption exceeds the one of As(+5). Note that only the OH groups that are associated with the Al ions exposed at the edges of clay particles, are considered to be proton acceptors and are able to complex anionic species of As [40,42]. In Table 3 we present selected geo-materials and also minerals, oxides and other constituents that can be use as low-cost effective absorbents for

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	Method /type of water	Optimum pH	Contamination concentration (mg/L)	Capacity (mg/g)		References
Adsorbent				As (III) As (V)		
Bentonite (modified)	Batch	6.0/9.0	0.2-1	0.82	1.48	[44]
Montmorillonite	Batch	6-May	20 µM	~0.2 mmol/g	~0.4 mmol/g	[42]
Illite	Batch	9	20 µM	~0.2 mmol/g	~0.5 mmol/g	[42]
Kaolinite	Batch	8-Mar	20 µM	~0.3 mmol/g	~0.5 mmol/g	[42]
Kaolinite, surfactant modified	Batch/column	5.0-6.5	0.2-14	4.3 mmol/kg	9.0 mmol/kg	[45]
Gibbsite	Wastewater	5.5	10-1000	3.3	4.6	[46]
Soil, Sharkey	Soil	5-6	5-100		0.74	[47]
Zeolite, surfactant modified	Batch/ column	7.2-7.5	0.2-14	1.6 mmol/kg	7.2 mmol/kg	[48,49]
Zeolites	Batch	4	0.1-4.0	0.017	0.1	[50]
Malachite	Batch	4	5		57.1 mg/g	[51]
Feldspar	Water/ wastewater	4.2	133.49 µmol/L		0.18	[52]
Siderite	Batch and column	7	250-2000	1040 µg/g	516 µg/g	[53,54]
Fe–Mn mineral material	Batch/column	3/3,5.5	0.47 mmol/L	14.7	6.7	[55]
Al <sub>2</sub> O <sub>3</sub> /Fe(OH) <sub>3</sub>	Batch	6.1 ± 0.3/8.0 ± 0.3	0.1-0.4	0.12 mmol/g	36.7	[56]
Fe–Mn binary oxide	Batch	4.8	0.20 mmol/L	1.77 mmol/g	0.93 mmol/g	[57,58]
Goethite	Batch wastewater	5.5	10-1000	7.5	12.5	[57,59]
Ferrihydrite	Batch/natural		325 µg/L		0.25	[60]
Ferric hydroxide, granular	Column drinking water	8-9	5-100	2.3		[61]
TiO <sub>2</sub>	Batch	8.5/7.3	0.4-80	32.4	41.4	[62,63]
Synthetic hydrotalcite	Ground water	7	400		105	[64]
Layered double hydroxides, calcined	Wastewater	4.2-5.4	20-200		5.61	[65]
FePO <sub>4</sub> (amorphous)	Drinking water	7-9/6-6.7	0.5-100	21	10	[66]
FePO <sub>4</sub> (cryst.)	Drinking water	7-9/6-6.7	0.5-100	16	9	[66]
Fe/NN-MCM-41	Drinking water	6	~0-1500		119.8	[67]
Cu/NN-MCM-48	Drinking water	7	~0-1500		37.46	[67]

Table 3: Constituents used for arsenic remediation purposes; adopted and modified from [43].

arsenic remediation purposes [43] (Figure 2). The suspension density is 40 g · L<sup>-1</sup>, and As total concentration is for the single-ion system 20  $\mu$ M and for the binary system As(+3)<sub>T</sub>=As(+5)<sub>T</sub>=20  $\mu$ M; adopted and modified from [44] (Table 3).

Redox fluctuations, pH and ions availability are responsible for the formation of insoluble sulfide precipitates such as arsenopyrite (FeAsS), realgar (AsS), and orpiment  $(As_2S_3)$  in reducing conditions [45]. Arsenic is also found in sedimentary environments, absorbed by Fe(+3) and Mn(+4) oxides – hydroxides after weathering of the sulfide minerals. In the case of the interaction of arsenopyrite with the ferric hydroxide, arsenic is adsorbed or co-precipitated with the ferrosoferric hydroxides via equation (19) [46]:

$$FeAsS + Fe(OH)_3 \rightarrow [Fe_x^{2+}Fe^{3+}(OH)_3^{x+}](AsO_4, AsO_3)_x + Products (19)$$

Therefore, high levels of arsenic in natural waters can be due to the reductive dissolution of arsenic rich iron oxyhydroxides [47,48]. In addition oxidative dissolution of arsenic-rich pyrite or arsenopyrite is responsible for As existence in natural waters [49]. In this case, the concentration of Dissolved Oxygen (DO) is the limiting factor for arsenopyrite dissolution in a variety of pH ranges [50,51]. Arsenic release rates seem to increase with increasing DO concentration and temperature, and are similar at low (<7) and high (>10) pH. The reaction includes FeAsS dissolution (equation 20) and As(+3) present as H<sub>3</sub>AsO<sub>3</sub>, while Fe(+2) is further oxidized to Fe(OH)<sub>3</sub> (equation 21), with an increase of acidity in the solution [52-58]:

$$4FeAsS + 11O_2 + 6H_2O \rightarrow 4Fe^{2+} + H_3AsO_3 + 4H_2SO_4^{2-}$$
(20)

$$4Fe^{2+} + O_2 + 10H_2O \to 4Fe(OH)_3 + 8H^+$$
(21)

The widespread arsenic contamination is thought to be related with As release from iron oxyhydroxides, probably due to the reaction of Fe-oxides/hydroxides with organic carbon [1,23,59-67]. In such a case the source of As is the adsorbed arsenic onto the surface of Fe-oxides/hydroxides solid phases, and a parallel release of arsenic during the reductive dissolution of ferric oxides-hydroxides occurs (equation 22) [68-70]:

$$FeOOH + CH_2O + 7H_2CO_3 \rightarrow 4Fe^{2+} + 8HCO_3^- + 6H_2O$$
 (22)

In addition, sorption of As(+3) onto Mn-oxides/hydroxides phases has been reported by Panagopoulos and Panagiotaras [71] in order to delineate controlling geochemical processes in the groundwater pool of the Trifilia karst aquifer, in Western Greece. The proposed mechanism is described by equation (23):

$$\equiv MnOH + As^{3+} \rightarrow \equiv MnOAs^{2+} + H^{+}$$
<sup>(23)</sup>

However, adsorbed arsenic species are weak acids and can affect the surface charge due to proton exchange reactions. Whether As adsorbs as a mononuclear or binuclear complex has implications for the level of protonation of the surface species, where this mechanism elucidated in equations (24) and (25) in the case of Fe-oxides/hydroxides surfaces [72-74]:

$$= FeOH + H_3AsO_3 \rightarrow = FeH_xAsO_3^{x-2} + H_2O + (2-x)H^+ \text{ where } x = 0-2$$
 (24)

$$(\equiv FeOH)_2 + H_3AsO_3 \rightarrow (\equiv Fe)_2H_yAsO_3^{y-1} + 2H_2O + (1-y)H^+ \text{ where } y = 0,1$$
(25)

Therefore, further reductive dissolution of ferric oxides-hydroxides by organic matter as described in equation (22), contributes to the cycling of arsenic species into the environment [16]. The overall



biogeochemical processes potentially influence arsenic mobility in the natural environment and actually generate specific patterns of distribution and speciation during redox and complexation reactions so that arsenic undergoes a variety of biogeochemical transformations.

# Conclusion

A variety of factors controls the fate of arsenic in the environment. The major biogeochemical transformations of arsenic constituents depended to the primary arsenic source, the redox potential, pH as well as microbial activity. However, the dissolution of arsenic minerals are the major source of the arsenic chemical species in the environment in specific physicochemical conditions. In different environmental conditions arsenic mineral degree of dissolution as well as its chemical species can be established using mathematical modeling.

These models can predict the occurrence of different arsenic chemical species throughout redox reactions taking place in specific temperature, pressure and pH conditions. Therefore, arsenopyrite oxidative dissolution is the major geochemical processes regulating the existence of arsenic in natural waters, with the dissolve oxygen concentration being responsible for the degree of dissolution in different pH values.

However, Fe-oxides/hydroxides solid phases adsorbed As and they can also release arsenic during their reductive dissolution. In aqueous systems, and in extremely low pH conditions (pH<2), arsenic acid occurs. In addition between pH=2 to pH=11, it is in the form of  $H_2AsO_4^-$  and  $HAsO_4^{-2}$  chemical species. As the pH increases from low values, and in mildly reducing conditions, arsenious acid is converting to  $H_2AsO_3^-$ .

Under oxic conditions Arsenate [As(+5)] is the thermodynamically stable phase of arsenic. In reducing conditions arsenite [As(+3)] ions occurring. The As(+5) chemical species are adsorbed onto hydrous ferric oxides and they also can be released during the microbial reduction of Fe(+3) solid phases.

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In addition, the mobility and fate of arsenic in the environment is also related to co-precipitation and adsorption onto clay minerals, manganese oxides and hydrous aluminum oxides. In the case of As(+5)ions, maximum adsorption capacities observed in low to up near 7 pH values for chlorite, illite, halloysite, kaolinite and montmorillonite clay minerals. However, adsorption capacity decreases as pH increases. An opposite behavior is apparent for the As(+3) regarding the same clay minerals. In this case adsorption of As(+3) increased with increasing pH, while adsorption was minimum at low pH values. As a concluding remark is that at s, As(+5) ions adsorbed to a greater extent than As(+3) on all the above mentioned clay minerals, while at higher pH values, adsorption of As(+5) and As(+3) were more comparable, and in some cases As(+3) adsorption exceeds that of As(+5). Therefore, clay minerals can be applied for arsenic removal and remediation purposes.

Although there is sufficient knowledge on the geochemical processes that governs the arsenic occurrence and fate in nature, the extent to how microbes affecting arsenic fluxes in the environment must be further studied in order to better understand the overall biogeochemical cycling of arsenic in the natural environment.

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#### References

- Smedley P, Kinniburgh D (2002) A review of the source, behaviour and distribution of arsenic in natural waters. Appl Geochem 17: 517-568.
- Edwards M, Johnson L, Mauer C, Barber R, Hall J (2014) Regional specific groundwater arsenic levels and neuropsychological functioning: a crosssectional study. Int J Environ Health Res 24: 546-557.
- Dewji N, Do C Bayney RM, (1995) Transcriptional activation of Alzheimer's β-amyloid precursor protein gene by stress. Mol Brain Res 33: 245–253.
- 4. Vahidnia A, van der Straaten RJHM, Romijn F, van Pelt J, van der Voet G (2007) Arsenic metabolites affect expression of the neurofilament and tau genes: an in vitro study into the mechanism of arsenic neurotoxicity. Tox In vitro 21: 1104-1112.
- Engström K, Vahter M, Johansson G (2010) Chronic exposure to cadmium and arsenic strongly influences concentrations of 8-oxo-7,8-dihydro-2'deoxyguanosine in urine. Free Rad Biol Med 48: 1211-1217.
- Vega L, Styblo M, Patterson R, Cullen W, Wang C (2001) Differential effects of trivalent and pentavalent arsenicals on cell proliferation and cytokine secretion in normal human epidermal keratinocytes. Tox and Appl Pharm 172: 225-232.
- Luo J, Qiu Z, Shu W, Zhang Y, Zhang L (2009) Effects of arsenic exposure from drinking water on spatial memory, ultra-structures and NMDAR gene expression of hippocampus in rats. Tox Lett 184: 121-125.
- Meng D, Wang X, Chang Q (2010) Arsenic promotes angiogenesis in vitro via a heme oxygenase-1-dependent mechanism. Tox Appl Pharm 244: 291-299.
- Tan Z, Seshadri S, Beiser A (2003) Plasma total cholesterol level as a risk factor for Alzheimer disease: the Framingham Study. Arch Int Med 163: 1053-1057.
- Chiou H, Hsueh Y, Liaw KF (1995) Incidence of internal cancers and ingested inorganic As: a seven-year follow-up study in Taiwan. Canc Res 55: 1296– 1300.
- 11. Mandal B, Suzuki K (2002) Arsenic round the world: A review. Talanta 58: 201-235.

- 12. U.S. EPA (2002) Arsenic Treatment Technologies for Soil, Waste, and Water. EPA-542-R-02 004.
- Mandal BK, Chowdhury TR, Samanta G, Mukherjee DP, Chanda CR (1998) Impact of safe water for drinking and cooking on five arsenic-affected families for 2 years in West Bengal, India. Sci Total Environ 218: 185-201.
- 14. Acharyya SK, Chakraborty P, Lahiri S, Raymahashay BC, Guha S (1999) Arsenic poisoning in the Ganges delta. Nature 401: 545-547.
- Nickson R, McArthur J, Burgess W, Ahmed KM, Ravenscroft P (1998) Arsenic poisoning of Bangladesh groundwater. Nature 395: 338.
- Nickson R, McArthur J, Ravenscroft P, Burgess W, Ahmed K (2000) Mechanism of arsenic release to groundwater, Bangladesh and West Bengal. Appl Geochem 15: 403-413.
- 17. Kumaresan M, Riyazuddin P (2001) Overview of speciation chemistry of arsenic. Curr Sci 80: 837-846.
- The Agency for Toxic Substances and Disease Registry (ATSDR): ToxFAQs<sup>™</sup> for Arsenic (2001) July 12.
- 19. Kirk-Othmer (1992) Arsenic and Arsenic Alloys. The Kirk-Othemer Encyclopedia of Chemical Technology, Volume 3. John Wiley and Sons, New York.
- 20. National Research Council (1999) Arsenic in Drinking Water. Washington, D.C. National Academy Press.
- Korte N (1991) Naturally occurring arsenic in groundwaters of the Midwestern United States. Environ Geol Water 18: 137-141.
- IPCS (2001) Environmental health criteria on arsenic and arsenic compounds. In Environmental Health Criteria Series: Arsenic and arsenic compounds, WHO, Geneva 224: 521.
- Welch A, Westjohn DB, Helsel DR, Wanty RB (2000) Arsenic in ground water of the United States: Occurrence and geochemistry. Ground Wat 38: 589-604.
- 24. Sarkar B (2002) Heavy metals in the environment. Marcell Dekker, New York.
- 25. Liao C, Chen B, Singh S, Lin M, Liu C (2003) Acute Toxicity and Bioaccumulation of Arsenic in Tilapia (Oreochromis mossambicus) from a Blackfoot Disease Area in Taiwan. Env Tox 18: 252-259.
- 26. Smith E, Smith J, Biswas T, Correll R, Naidu R (2003) Arsenic in Australian Environment: An Overview. J Env Sci and Health, Part A: Toxic/Hazardous Substances and Environ Eng 38: 223-239.
- Panagiotaras D, Papoulis D, Stathatos E (2015) Origin of Arsenic Toxicity Geochemistry. Chapter 4, in Arsenic Toxicity: Prevention and Treatment. Editor: Narayan Chakrabarty, CRC press, Taylor & Francis publishers.
- Azcue JM, Mudroch A, Rosa F, Hall GEM, Jackson TA (1995) Trace elements in water, sediments, porewater, and biota polluted by tailings from an abandoned gold mine in British Columbia, Canada. J Geochem Expl 52: 25-34.
- U.S. EPA (2000) National Primary Drinking Water Regulations; Arsenic and Clarifications to Compliance and New Source Contaminants Monitoring; Proposed Rule. Federal Register 65, 121: 38888.
- Drahota P, Filippi M (2009) Secondary arsenic minerals in the environment: A review. Env Inter 35: 1243-1255.
- Nordstrom DK, Archer DG (2003) Arsenic thermodynamic data and environmental geochemistry. In: Welch AH, Stollenwerk KG, editors. Arsenic in ground water 1-25. Boston: Kluwer Academic Publishers.
- 32. Cullen WR, Reimer KJ (1989) Arsenic speciation in the environment. Chem Rev 89: 713-764.
- Dowdle Philip R, Laverman Anniet M, Oremland Ronald S (1996) "Bacterial reduction of arsenic(V) to arsenic(III) in anoxic sediments." Appl Environ Microbiol 62: 1664-1669.
- Lièvremont D, Bertin PN, Lett MC (2009) Arsenic in contaminated waters: Biogeochemical cycle, microbial metabolism and biotreatment processes. Biochimie 91: 1229-1237.
- Rakhunde R Jasudkar D Deshpande L Juneja HD Labhasetwar P (2012) Health Effects and Significance of Arsenic Speciation in Water. Inter J Env Sci Res 1: 92-96.
- Tolm TR, Kelly WR, Wilson SD, Roadcap GS, Talbott JL (2004) Arsenic Geochemistry and Distribution in the Mahomet Aquifer, Illinois. Illinois Waste

Management and Research Center. Illinois Department of Natural Resources 1-103.

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- 37. Stumm W, Morgan J (1996) Aquatic Chemistry: Chemical Equilibria and Rates in Natural Waters. New York: John Wiley and Sons.
- 38. Oremland RS, Stolz JF (2003) The ecology of arsenic. Science 300: 939-944.
- 39. Paez-Espino D, Tamames J, De Lorenzo V, Cánovas D (2009) Microbial responses to environmental arsenic. Biol Met 22: 117-130.
- Stollenwerk KG (2003) Geochemical processes controlling transport of arsenic in groundwater: A review of adsorption. In:. Arsenic in ground water, Welch AH, Stollenwerk KG, Boston: Kluwer Academic Publishers.
- Zheng C, Bennett GD (2002) Applied Contaminant Transport Modeling, Theory and Practice. New York, Van Nostrand Reinhold.
- Goldberg S (2002) Competitive Adsorption of Arsenate and Arsenite on Oxides and Clay Minerals. Soil Sci Soc Am J 66: 413-421.
- 43. Papoulis D, Panagiotaras D, Panagopoulos G (2012) Arsenic: Environmental Impact Reduction Using Natural and Modified Adsorbents. In Arsenic: Sources, Environmental Impact, Toxicity and Human Health - A Medical Geology Perspective. NOVA Publishers 289-304.
- 44. Su J, Huang H-G, Jin X-Y, Lu X-Q, Chen Z-L (2011) Synthesis, characterization and kinetic of a surfactant-modified bentonite used to remove As(III) and As(V) from aqueous solution. J Hazard Mater 185: 63-70.
- 45. Matsunaga H, Yokoyama T, Eldridge RJ, Bolto BA (1996) Adsorption Characteristics of Arsenic(III) and Arsenic(V) on Iron(III)-Loaded Chelating Resin Having Lysine-Nα, Nα -Diacetic Acid Moiety. React Funct Polym 29: 167-174.
- Ladeira ACQ, Ciminelli VST (2004) Adsorption and desorption of arsenic on an oxisol and its constituents. Water Res 38: 2087-2094.
- Zhang H, Selim HM (2005) Kinetics of arsenate adsorption-desorption in soils. Environ Sci Technol 39: 6101-6108.
- Li Z, Beachner R, McManama Z, Hanlie H (2007) Sorption of arsenic by surfactant-modified zeolite and kaolinite. Micropor Mesopor Mater 105: 291-297.
- Sullivan EJ, Bowman RS, Legiec IA (2003) Sorption of arsenic from soilwashing leachate by surfactant-modified zeolite. J Environ Qual 32: 2387-2391.
- Elizalde-Gonzalez MP, Mattusch J, Einicke WD, Wennrich R (2001) Sorption on natural solids for arsenic removal. Chem Eng J (Lausanne) 81: 187-195.
- Saikia J, Saha B, Das G (2011) Efficient removal of chromate and arsenate from individual and mixed system by malachite nanoparticles. J Hazard Mater 186: 575-582.
- Singh DB, Prasad G, Rupainwar DC (1996) Adsorption technique for the treatment of As (V)-rich effluents. Colloid Surf A 111: 49–56.
- 53. Sajidu SMI, Persson I, Masamba WRL, Henry EMT, Kayambazinthu D (2006) Removal of Cd<sup>2+</sup>, Cr<sup>3+</sup>, Cu<sup>2+</sup>, Hg<sup>2+</sup>, Pb<sup>2+</sup> and Zn<sup>2+</sup> cations and AsO<sub>4</sub><sup>3-</sup> anions from aqueous solutions by mixed clay from Tundulu in Malawi and characterisation of the clay. Water SA 32: 519-526.
- Maji SK, Pal A, Pal T (2007) Arsenic removal from aqueous solutions by adsorption on laterite soil. J Environ Sci Health A 42: 453-462.
- Deschamps E, Ciminelli VS, Höll WH (2005) Removal of As (III) and As(V) from water using a natural Fe and Mn enriched sample. Water Res 39: 5212-5220.
- Hlavay J, Polyak K (2005) Determination of surface properties of iron hydroxidecoated alumina adsorbent prepared for removal of arsenic from drinking water. J Colloid Interface Sci 284: 71-77.
- Zhang G, Qu J, Liu H, Liu R, Wu R (2007) Preparation and evaluation of a novel Fe–Mn binary oxide adsorbent for effective arsenite removal. Water Res 41: 1921-1928.
- Gao SZ, Jiu HQ, Hul JL, Rui PL, Guo TL (2007) Removal Mechanism of As(III) by a Novel Fe–Mn Binary Oxide Adsorbent: Oxidation and Sorption. Environ Sci Technol 41: 4613-4619.
- 59. Ladeira ACQ, Ciminelli VST (2004) Adsorption and desorption of arsenic on an oxisol and its constituents. Water Res 38: 2087-2094.
- 60. Tobias R, Jordi C, Carlos A, Cortina JL, De Pablo J (2006) Use of caustic

magnesia to remove cadmium, nickel, and cobalt from water in passive treatment systems: column experiments. Environ Sci Technol 40: 6438-6443.

- 61. Daus B, Wennrich R, Weiss H (2004) Sorption materials for arsenic removal from water: a comparative study. Water Res 38: 2948-2954.
- Zhang FS, Itoh H (2006), Photocatalytic oxidation and removal of arsenite from water using slag-iron oxide-TiO<sub>2</sub> adsorbent. Chemosphere 65: 125-131.
- Bang S, Patel M, Lippicott L, Meng XG (2005) Removal of arsenic from groundwater by granular titanium dioxide adsorbent. Chemosphere 60: 389-397.
- 64. Kiso Y, Jung YJ, Yamada T, Nagai M, Min KS (2005) Removal properties of arsenic compounds with synthetic hydrotalcite compounds. Water Sci Technol 5: 75-81.
- 65. Yang L, Shahrivari Z, Liu PKT, Sahimi M, Tsotsis TT (2005) Removal of Trace Levels of Arsenic and Selenium from Aqueous Solutions by Calcined and Uncalcined Layered Double Hydroxides (LDH). Ind Eng Chem Res 44: 6804-6815.
- Lenoble V, Laclautre C, Deluchat V, Serpaud B, Bollinger JC (2005) Arsenic removal by adsorption on iron(III) phosphate. J Hazad Mater 123: 262-268.
- 67. Yoshitake H, Yokoi T, Tatsumi T (2003) Adsorption Behavior of Arsenate

at Transition Metal Cations Captured by Amino-Functionalized Mesoporous Silicas. Chem Mater 15: 1713-1721.

- Kim MJ, Nriagu J, Haack S (2002) Arsenic species and chemistry in groundwater of southeast Michigan. Environ Poll 120: 379-390.
- 69. Yu Y, Zhu Y, Gao Z, Gammons CH, Li D (2007) Rates of arsenopyrite oxidation by oxygen and Fe(III) at pH 1.8–12.6 and 15–45°C. Environ Sci and Tech 41: 6460-6464.
- 70. Corkhill CL, Vaughan DJ (2009) Arsenopyrite oxidation-A review. Appl Geochem 24: 2342-2361.
- Panagopoulos G, Panagiotaras D (2011) Understanding the extent of geochemical and hydrochemical processes in coastal karst aquifers through ion chemistry and multivariate statistical analysis. Fres Environ Bull 20: 3270-3285.
- Sverjensky DA, Fukushi K (2006) Anion adsorption on oxide surfaces: inclusion of the water dipole in modeling the electrostatics of ligand exchange. Environ Sci and Techn 40: 263-271.
- 73. Jain A, Loeppert RH (2000) Effect of competing anions on the adsorption of arsenate and arsenite by ferrihydrite. J Envir Qual 29: 1422-1430.
- Goldberg S, Johnston CT (2001) Mechanisms of Arsenic Adsorption on Amorphous Oxides Evaluated Using Macroscopic Measurements, Vibrational Spectroscopy, and Surface Complexation Modeling. J Coll Inter Sci 234: 204-216.

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