

A Point of Care Real Time PCR Platform Based on Silicon Technology

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Abstract

The miniaturization of a real time PCR amplification systems are key to genetic point-of-care (POC) diagnostics to offer *sample-in answer-out* analysis. Actually, miniaturization typically enables shorter analysis times, reduces reagent consumption, minimizes risk of sample contamination, and often enhances assay performance. Here we present the genetic real time PCR PoC system based on silicon microchip able to address the quantitative and qualitative identification of multiple analytes of specific nucleic acids (DNA, RNA) sequences. Thanks to the combination of integrated silicon temperature sensors and heaters, the specific chip design architecture and the smart detection software, an improvement of sensitivity is achieved respect to the commercial tools.

Keywords: Real time PCR; Microchip; DNA

Introduction

The real-time PCR (Polymerase Chain Reaction) methodology has become an indispensable tool in many fields of molecular diagnostics, including determination of viral or bacterial loads in clinical samples, identification and titers of germs in food, diagnosis of tumors, gene expression analysis, or forensic analyses [1-3]. It is a wide used methodology since it is able to provide in a closed-system the quantitative result of target DNA sequence as it accumulates in real time in the reaction vessel. That allows avoiding time-consuming post-PCR analysis and reduces the risk of cross-contaminations enhancing both accuracy and speed.

The miniaturization of real time PCR amplification systems is a key point towards the development of the so called “genetic point-of-care (PoC)” that are able to offer *sample-in answer-out* diagnostic analysis [4,5]. Several solutions based on multiple systems to integrate the sample preparation, nucleic acids amplification and detection via real time PCR [6], LAMP [7] or microarray [8] has been reported in the literature. All these miniaturized devices have been designed to enable shorter analysis times, reduce reagent consumption, minimize risk of sample contamination and enhance the assay performance such as sensitivity, selectivity and limit of detection.

In this context, silicon is very attractive material for biochips due to both its physical aspects for sensing (such as, the low heat capacity, the good thermal conductivity and the possibility to be made porous to increase the surface-area and reaction efficiency) and its technological aspects (consolidated production technologies and industrialization processes, integration of electrodes to the chip, as well as “intelligence on board” with microelectronics circuitry, high-volume production). Additionally, the coming of MEMS (Micro-Electro Mechanical System) technology has open the perspective to integrate micropumps, micronozzle and sensor combining multiple functions to achieve a complete analysis solution, including fluid management, amplification, hybridization or affinity binding, and detection.

For an optically transduced platform the implementation of software for the image analysis is also crucial to achieve the signal improvement. In particular for an optimized fluorescence analysis, it is extremely important to precisely detect the region of interest (ROI) areas where the PCR takes place. The alignment between the chip reaction chambers and the ROI areas of the analysis is therefore fundamental. When a fixed selection of ROI areas is employed, errors in reading fluorescence are generated. In order to solve this

problem has been implemented a new flow process, integrating some different algorithm and creating smart decision system which detect automatically ROI areas, computing average values on that areas and calculate PCR for every cycle [9-12].

Here we present the miniaturized Bio2Bit Q3 Real time PCR platform able to perform quantitative and qualitative identification of multiple analytes of specific nucleic acids (DNA, RNA) sequences in a PoC format. The platform is based on silicon microchip that integrates in the same device both real time PCR micro reactors (6-15 μ l in volume) and silicon temperature sensors and heaters. Thanks to the integrated silicon temperature sensors and heaters, the chip allows a temperature control accuracy of $\pm 0.2^\circ\text{C}$, heating rate of 15°C/s and cooling rate of 8°C/s . Additionally, a specific chip design architecture [13] enhances the optical fluorescent signal in combination with a smart detection software, so that an improvement of sensitivity is achieved respect to the commercial tools. The chip is thermally and optically driven by a miniaturized instrument specifically developed for it design.

Here we report real time PCR applicative results related to the detection of hepatitis B virus (HBV) using a specific HBV clone as sample material.

Materials and Methods

qRT-PCR platform

The real time PCR System is composed by the following components: (a) a disposable chip that contains reaction chambers for the real time PCR process; (b) a portable and customized thermocycler instrument (Q3 reader) that thermally and optically drives the chip during the real time PCR process; (c) a software package that manages the PCR process and carried out the data analysis.

Figure 1 reports the picture of the platform.

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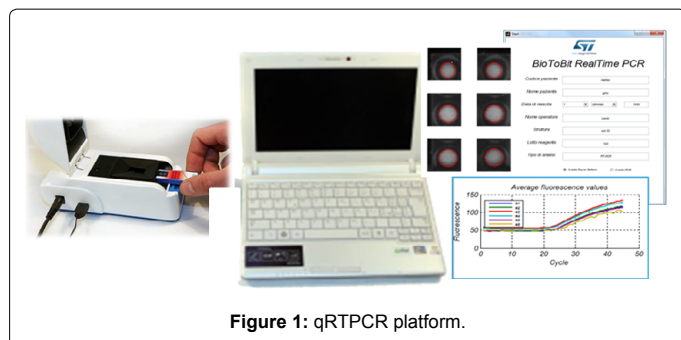


Figure 1: qRT-PCR platform.

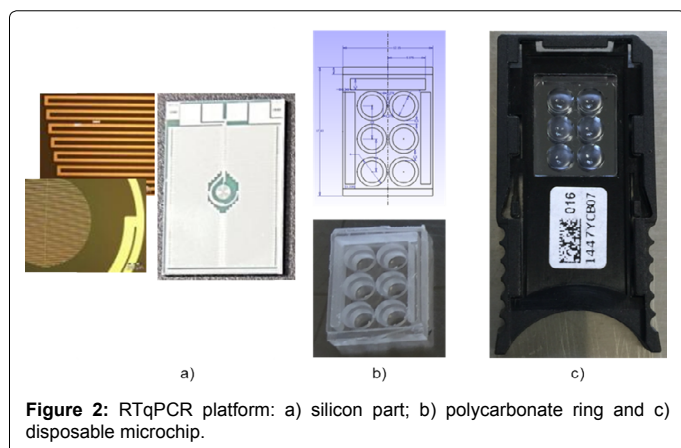


Figure 2: RTqPCR platform: a) silicon part; b) polycarbonate ring and c) disposable microchip.

Microchip fabrication

The core of the platform is a silicon-plastic hybrid microchip (Figure 2) composed by:

(a) A bottom part made in silicon by the VLSI (Very-large-scale integration) technology on a six inch silicon wafer that contains the temperature sensors and heaters integrated on the bottom of the device (Figure 2a). They consist in AlCu metal strips (size 4.0 ± 0.1 , spaced $5.0 \pm 0.1 \mu\text{m}$ for the temperature sensor, size $115.0 \pm 0.1 \mu\text{m}$ spaced $30.0 \pm 0.1 \mu\text{m}$ for the heater). The device perform thermal cycling in combination with the driving Instrument (Q3-reader below described) with the following performances: temperature sensor maximum power consumption 55 mW, heater maximum power consumption 10 mW, temperature control accuracy of $\pm 0.2^\circ\text{C}$, heating rate of $15^\circ\text{C}/\text{s}$, cooling of rate: $8^\circ\text{C}/\text{s}$ and temperature resolution: 0.1°C .

(b) A top part (ring) made in polycarbonate. The ring was manufactured by molding technology and comprises an architecture containing 6 reaction chambers with cylindrical shape featured by a volume of 25 μL (Figure 2b). The ring is glued onto the silicon part by silicone glue. The microchip is finally mounted on a plastic holder to facilitate the chip handling (Figure 2c).

During the PCR cycles the thermal sealing is guaranteed by 15 μL of paraffin wax dispensed on each chamber.

Q3 Reade

The Q3 instrument reader was developed by STMicroelectronics (Figure 1). It thermally and optically drives the chip during the real time PCR process. It has a size of $14 \text{ L} \times 7.1 \text{ W} \times 8.7 \text{ H}$ (cm) (weight: 322 g). The optical module contains up to 2 independent optical channels, for multiple Fluorescent reporters (FAM, VIC[®]). The LED

light sources are centered at wavelength 470, 530 nm, respectively. The emission is detected by a CMOS camera that contains a high-pass filter at 520 nm or 20 nm wide band-pass filter centered at 556. Finally, the thermal Module drives the temperature sensor and heater of the chip to reach the temperature performance characteristics described in the section 2.2.

Smart-software for image analysis

The smart-software was properly developed for the image analysis, it is based on three main core modules: 1) Fully automatic image quantification: a set of algorithms for the digital image analysis process; 2) Numerical computation: a set of algorithms for the numerical analysis of raw data coming from real time PCR, 3) Graphical output: a graphical user interface to show the fluorescence curves for the PCR.

Chemicals

Hepatitis B virus (HBV) clone (ref. product CLO-05960116 HBV Complete Genome) and all the reagents for the HBV Real time PCR were purchased from Clonit (kit ref. product CLO-FO2 HBV MMIX KIT 48) and used according to the instruction for use.

RT-PCR experimental set up

Real time PCR experiments were performed using a Master Mix solution (volume 10 μL) containing Clonit buffer (1 \times) and Taq DNA polymerase, 0.5 μM of each forward and reverse primers, 2 μL of HBV clone (10^5 copies/ μL). Different amount of probe (0.01, 0.05, 0.1 and 0.2 μM) were tested. The PCR cycling was performed in the portable Q3-thermocycler-reader by using the following thermal program: preheating period of 10 min. at 95°C , followed by a maximum of 45 cycles of 95°C for 15s and 60°C for 60s. Different temperature of annealing were used for testing (60°C - 63°C). Negative samples were prepared substituting the HBV clone with the same amount of water. Each experiment was repeated 10 times.

Same real time PCR experiments (10 μL of the above reported Master Mix) were executed on standard 96 well reaction plate in gold standard Applied Biosystem 7500 real time PCR equipment as reference.

Results and Discussion

In order to demonstrate the ability of the system to detect the pathogen species HBV, we conduct real time PCR experiments with different conditions. In particular in order to optimize the device performance, we explored different annealing temperatures and probe concentrations. Figure 3a and 3b reports a typical qRT-PCR curves and fluorescence images obtained during the above described experiments

More in details, an amount of 15 μL of PCR master mix (containing HBV-clone 10^5 cps/ μL) was loaded on each chamber (the loading process was performed at 70°C in order to melt the paraffin wax) and the amplification at three different annealing temperatures (60, 62 and 63°C) was conducted. The fluorescence images after each cycles was analyzed by the smart-software and the Ct values calculated. The results are reported in Figure 4a. The data indicate no relevant difference in term of Ct for the three temperatures investigated with an average value for Ct of about 21.5. The reference samples processed on gold standard 96-well reaction plate in Applied Biosystem 7500 equipment report for the testing at different annealing temperature an average value of $\text{Ct } 22.4 \pm 0.5$. These data indicate an improvement on PCR efficiency performed on microchip device of about 1 Ct respect to the commercial equipment. This advance was explained by the specific

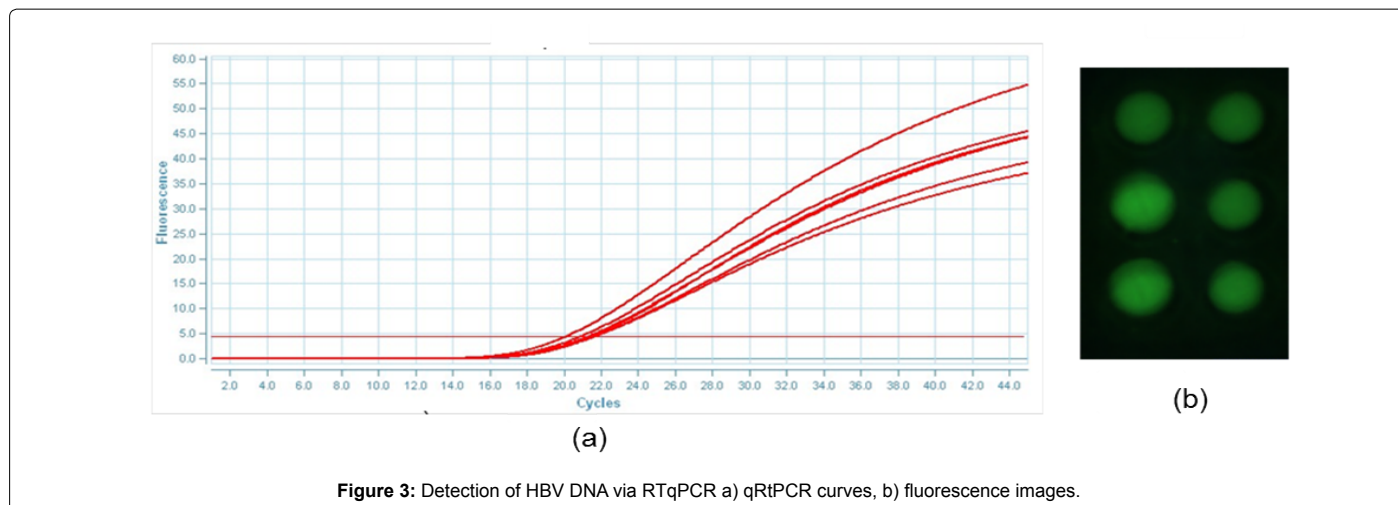


Figure 3: Detection of HBV DNA via RTqPCR a) qRtPCR curves, b) fluorescence images.

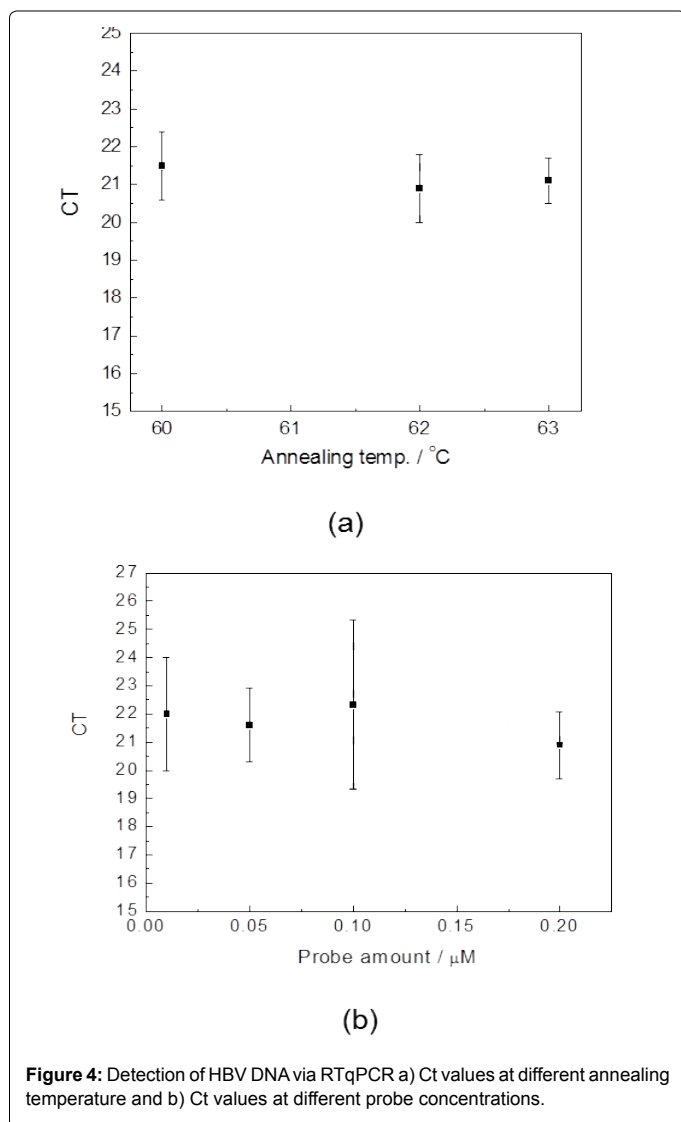


Figure 4: Detection of HBV DNA via RTqPCR a) Ct values at different annealing temperature and b) Ct values at different probe concentrations.

microchip design architecture that include a chamber designed to collect the paraffin wax (15 μ L) on the upper cylindrical part (diameter 3.5 mm) while the biological mix (10 μ L) is collected on the lower zone

(diameter 3.2 mm): this shape in combination with a smart detection software improve the final signal enhances the optical fluorescent signal of platform.

In order to investigate the robustness of the assay at the annealing temperature of 60°C (since no difference in the annealing temperature was found-*vide supra*), additional experiments were performed with different probe amounts. With the same method above described, an amount of 15 μ L of PCR of master mixes containing a concentration of 10⁵ cps/ μ L HBV-clone and different probe amount (0.01, 0.05, 0.1 and 0.2 μ M), were loaded into the microchip pre-heated chambers and real time PCR amplifications carried out. Figure 4b reports the related experimental results. Also in this case no variation of Ct was found with an average value for Ct of about 21.3. No fluorescence signal was registered for the negative sample (data not showed). The reference samples processed on gold standard 96-well Applied Biosystem 7500 show an average value of Ct 22.7 \pm 0.4. These results proves the assay is robust in the experimental condition ranges confirming the improvement of sensitivity of about 1 Ct compared to the commercial equipment. Additionally, they indicate that the platform allows to use very low amount of probe (0.01 μ M) without any efficiency variation.

Conclusions

Here we presented a miniaturized platform for nucleic acids detection by Real time PCR. The core of the platform is a hybrid microchip composed by a silicon part and a polycarbonate portion mounted on the plastic ring. Each PCR chamber with a total volume of volume 25 μ L is pre-loaded with 15 μ L of paraffin wax to guarantee the sealing during the thermal protocol. The chamber has a particular shape suitable to accommodate an amount of 10 μ L of biological master mix. The experiments were conducted at different annealing temperature and probe amounts and the results here discussed indicate an improvement of sensitivity of about 1 Ct, thanks to the smart-software implemented for the image analysis and to the chamber shape.

Acknowledgments

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